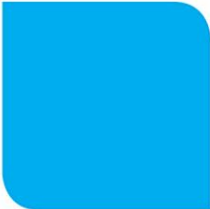


Droplet Digital™ PCR Training

Charlene Lu



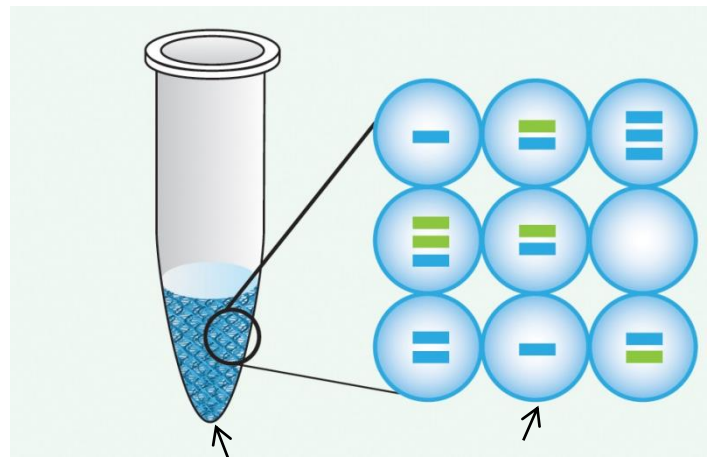
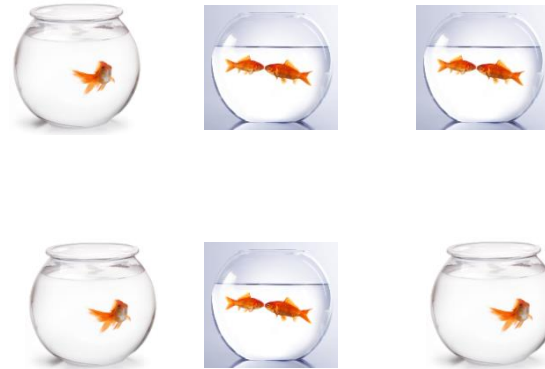
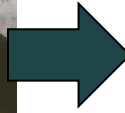


Outline

- What is digital PCR?
- What's Bio-Rad digital PCR?
- How is Droplet Digital PCR (ddPCR™) performed on the QX200™ Droplet Digital PCR systems
- Applications updated based upon QX100/QX200 Published data
 - Copy Number Variation
 - Rare Event Mutation detection
 - Gene Expression
- Summary

What is Digital PCR?

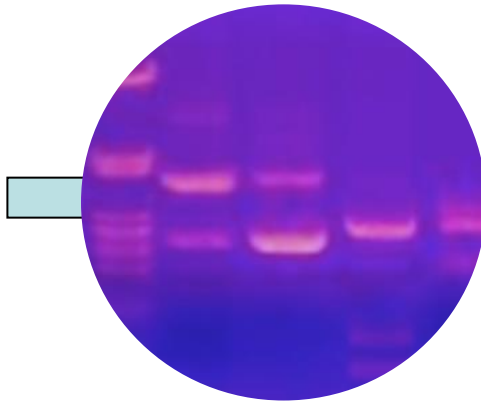
PCR reaction that is partitioned.



Many thousands
of discrete measurements

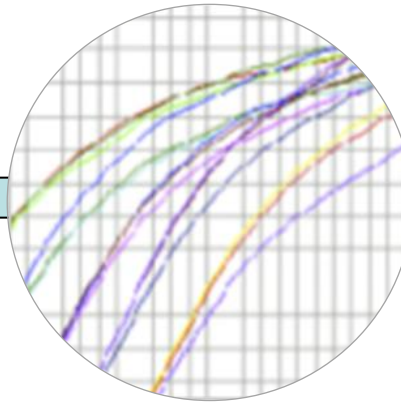
Generations of PCR

1st Generation



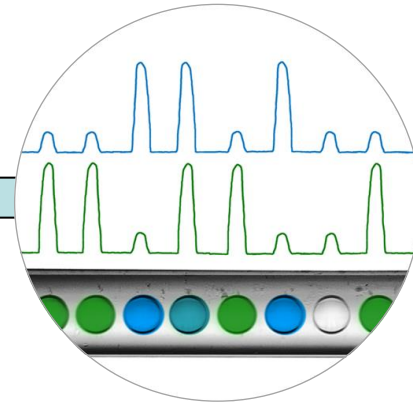
Gel Electrophoresis
(qualitative)

2nd Generation



Real-Time PCR
(indirect quantification)

3rd Generation

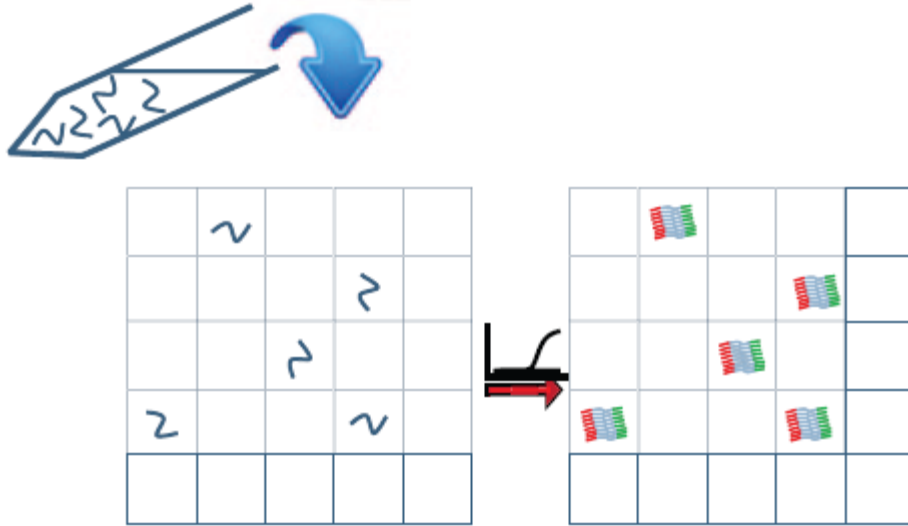


Droplet Digital PCR
(absolute quantification)

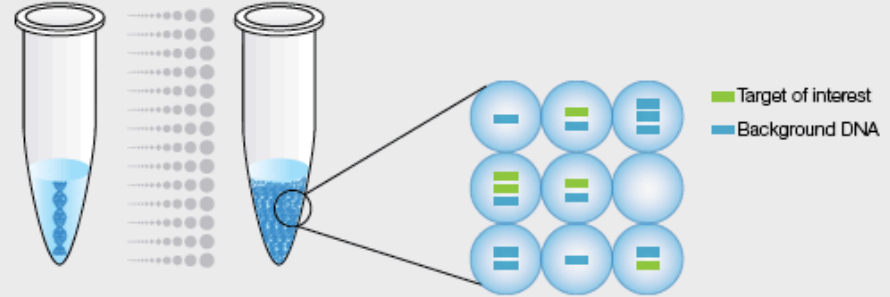
- End point (0's or 1's)
- Less sensitive to PCR efficiency
- No standard curve
- More tolerant to PCR inhibitors

ddPCR improves precision, sensitivity and reproducibility

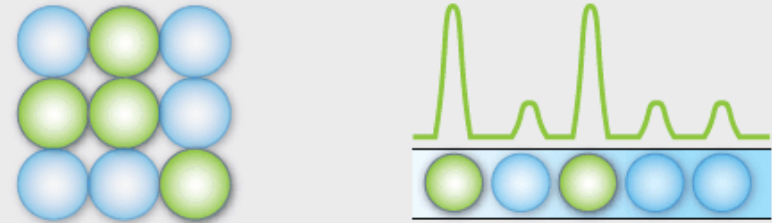
dPCR Principle



- End point (0's or 1's)
- Less sensitive to PCR efficiency
- No standard curve
- More tolerant to PCR inhibitors



The sample is partitioned into 20,000 droplets, with target and background DNA randomly distributed among the droplets.

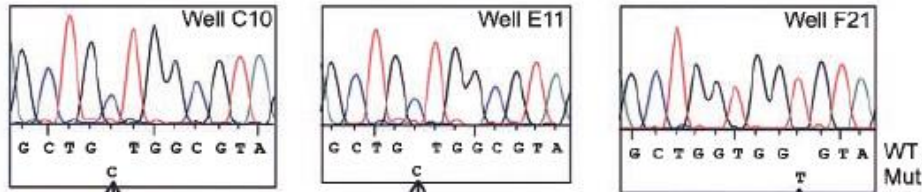


After PCR amplification, each droplet provides a fluorescent positive or negative signal indicating the target DNA was present or not present after partitioning. Each droplet provides an independent digital measurement.

“X” target copies

Positive and negative droplets are counted for the sample and the software calculates the concentration of target DNA as copies per microliter.

The 1st Paper about dPCR



- Mutant allele (KRAS)
- Wild type allele
- No amplification
- Positive control
- Negative control



Proc. Natl. Acad. Sci. USA
Vol. 96, pp. 9236–9241, August 1999
Genetics

Digital PCR

BERT VOGELSTEIN* AND KENNETH W. KINZLER

The Howard Hughes Medical Institute and the Johns Hopkins Oncology Center, Baltimore, MD 21231

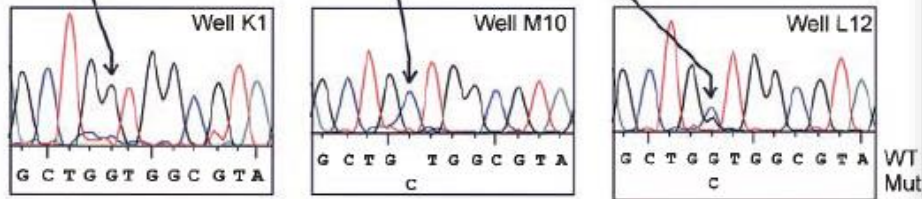
Contributed by Bert Vogelstein, June 9, 1999

Bert Vogelstein (born 1949) is a [Howard Hughes Medical Institute](#) investigator^[1] at The [Johns Hopkins University](#). He clarified the role of the gene [p53](#), which repairs DNA in dividing cells and destroys the cell if its DNA cannot be repaired. Damaged p53 is responsible for half of all cancers. More recently, his group sequenced the DNA of human breast and colon cancer, identifying genes which are mutated in each cancer.

Vogelstein developed the concept that some genes, such as TP53, KRAS, and APC are involved in cancer with great frequency, in close to 100% of some cancers; he called these genes "mountains." But thousands of genes are involved in cancer but are found at very low frequency, under 5%; he called these genes "hills." Collectively, however, the hills are also required for most cancers.

He found that while the number and complexity of these thousands of genes might be bewildering, most of them can be grouped into twelve critical pathways, such as [apoptosis](#), DNA damage control, invasion, [cell cycle](#) signaling, [KRAS](#) signaling, and [TGF-beta](#) signaling.

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24
A	0.9	0.8	0.7	1.1	0.8	0.8	0.9	0.9	0.9	0.8	1.0	0.9	1.1	0.9	1.0	0.9	1.0	0.8	0.9	1.0	1.0	1.0	0.9	0.9
B			1.3						0.9	0.8	1.0		1.1							1.0	1.0			
C	0.8								0.6	0.4		1.1	1.0	0.9	0.9									0.5
D				1.0					1.0	0.9	0.8	0.8				0.9						0.8	1.0	
E		1.0	1.0								0.7	0.9	0.9						0.8			1.1		
F				1.0			0.9	1.0			0.7		1.1							0.8	0.8	0.6	0.6	
G		0.9					0.9				0.8	0.8		1.0			1.0	0.9	1.1			1.0	1.0	1.2
H																								
I	1.1	1.1	1.2	1.1	0.9	1.1	1.1	0.9	1.2	1.1	1.2	1.1	1.0	1.2	1.0	1.0	1.0	1.1	1.0	1.0	1.0	1.0	1.2	1.4
J							1.0									0.9		1.2						
K	1.2	1.1	0.9						1.3	1.1	1.1							1.1		0.9			0.9	0.8
L				0.9	0.6							0.7	0.9					1.2	1.2				1.1	
M				0.7	0.9				0.8	1.0	0.4		0.7	0.9		1.1	1.1					1.1	1.1	
N			0.8	0.7				0.7	0.7		0.8					1.0					1.2		1.2	0.8
O					1.1			0.8			0.8		0.9								1.2			
P																								



How does Partitioning Enable Molecular Counting?

One Partition — One Reaction, One Data Point



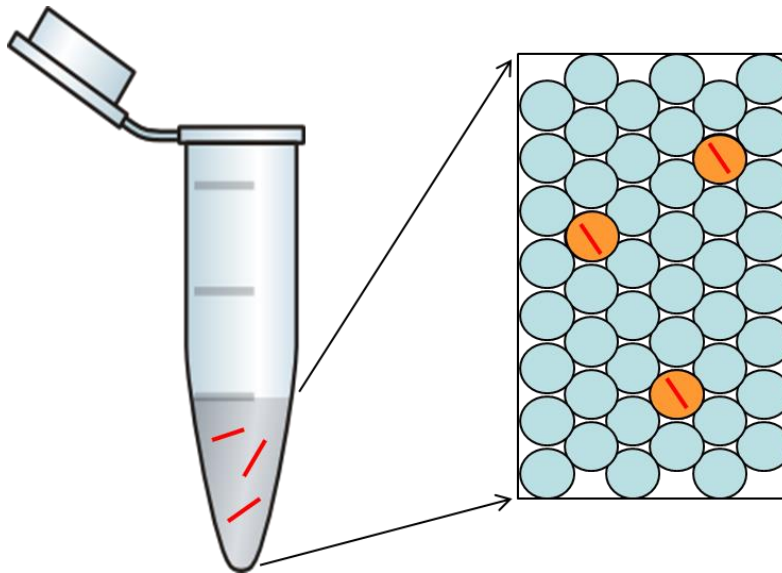
One measurement



Nanodroplet PCR reactions are independent, single amplification events



Many thousands of discrete measurements



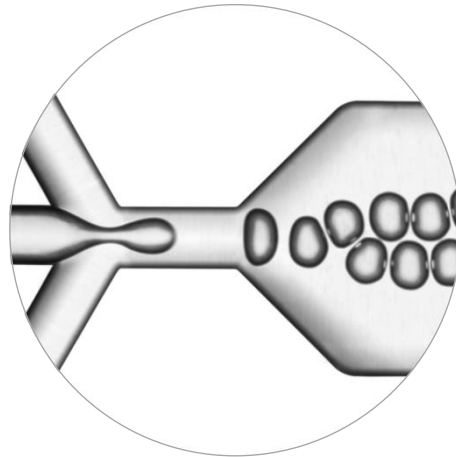
- Every partition is an individual reaction vessel
- End-point PCR
- If a partition has a target molecule it will be read as a positive
- If a partition has no target molecules it will be read as a negative
- In quantitative PCR (qPCR) — time course, quantification cycle (C_q) or threshold cycle (C_T), standard curves



Droplet Digital PCR Work Flow

Workflow of Droplet Digital PCR

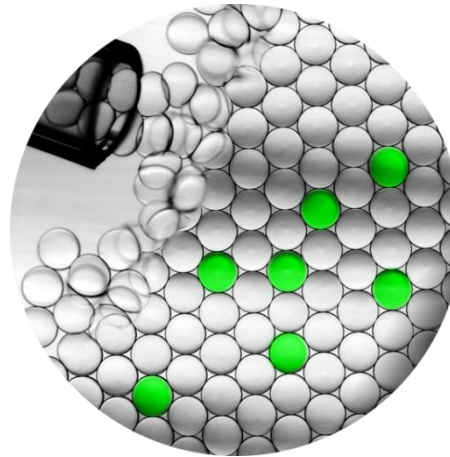
1. Make Droplets



Droplet Generator



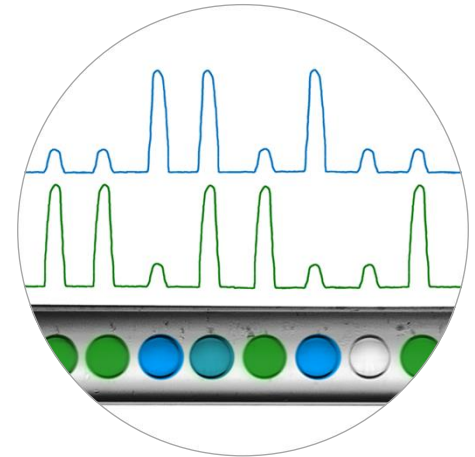
2. Cycle Droplets



Bulk PCR
Thermal Cycler



3. Read Droplets



Droplet Reader

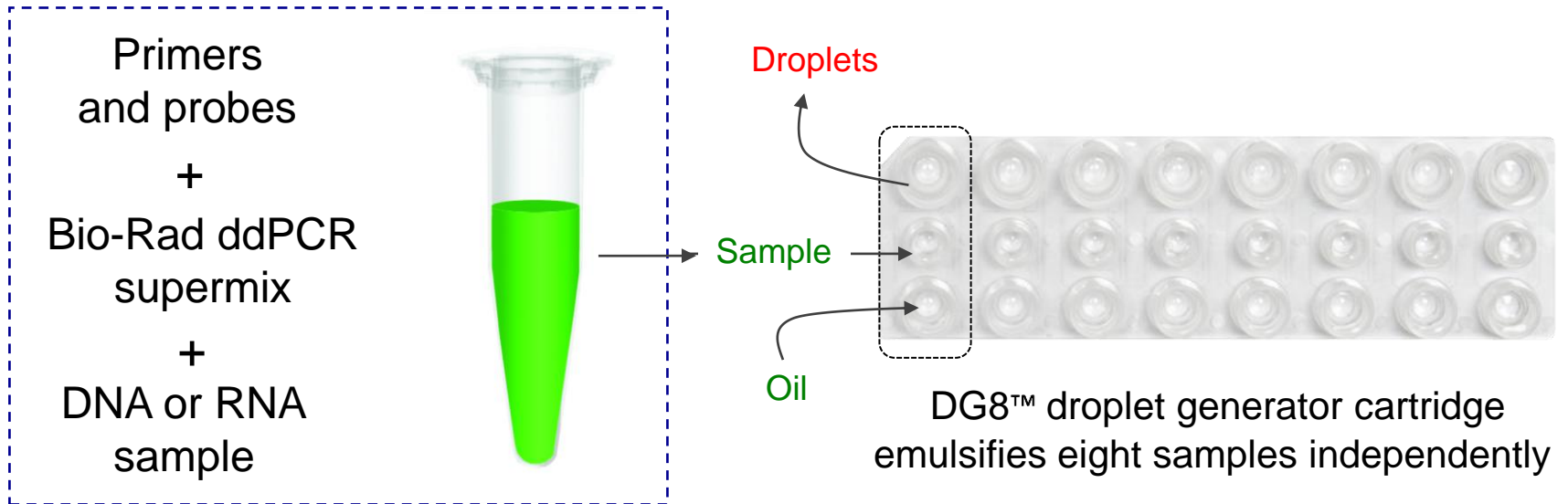


Primers and
optional probes
ddPCR supermix
sample

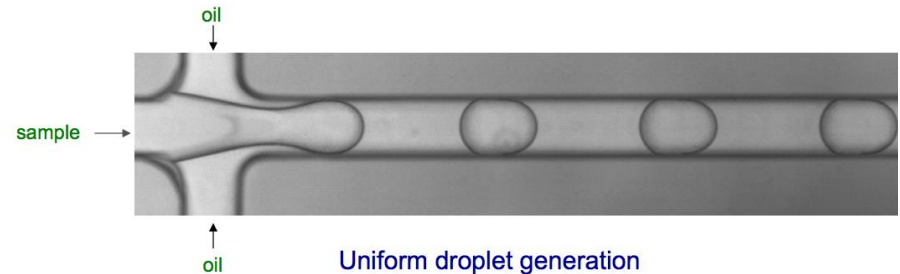
- Readout: copies/ μ l
- Dynamic range: 1–100,000 copies/well (~330 ng human genomic DNA)

Prepare Sample and Reagent Mixture

Prepare samples **exactly the same** as qPCR or PCR



Compatible with probes (FAM and HEX/VIC) or EvaGreen

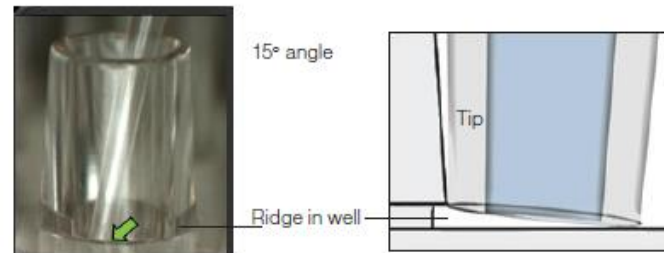


1. Insert DG8 cartridge and Add Sample



- ✓ All 8 sample wells must contain sample or 1x control buffer and 8 oil wells must contain droplet generation oil.
- ✓ Add 20ul sample before add oil
- ✓ Avoid bubbles

Inserting the DG8 cartridge into the cartridge holder.



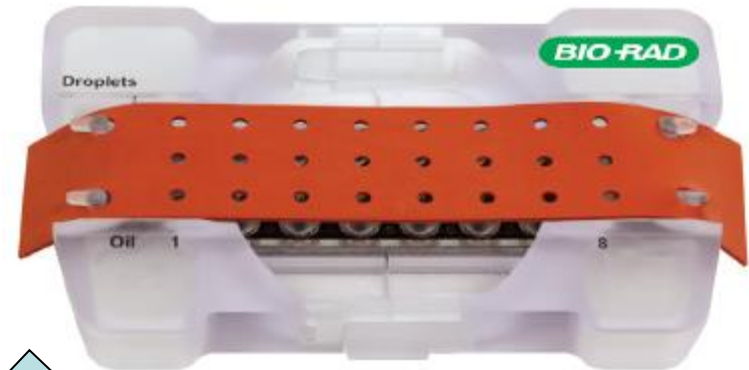
Transferring sample to the sample wells (middle row) of the DG8 cartridge. Hold the pipet tip at a 15° angle and at the bottom of the well (middle and right panels); do not dispense sample onto the wall or side of the well.

2. Add oil and place the gasket

- ✓ Add 70ul generation oil (check oil type)



- ✓ Hook the gasket



Press to open and close

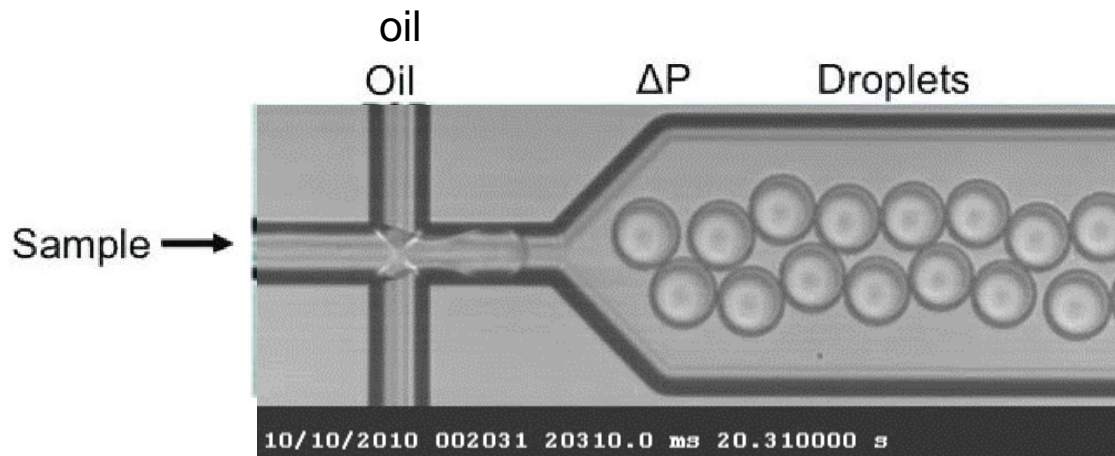
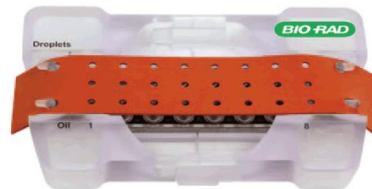


Status indicator lights



Droplet Generation by Flow Focusing

- Place loaded cartridge into QX200 Droplet Generator
- Generate 20,000 droplets per sample, 2 ½ min for 8 samples
- Average droplet size is 0.91 nl volume (125 micron diameter)



Uniform droplet generation

3. Transfer the droplets and seal the plate



Aspirating droplets from the DG8 cartridge.



Dispensing droplets into a 96-well PCR plate.

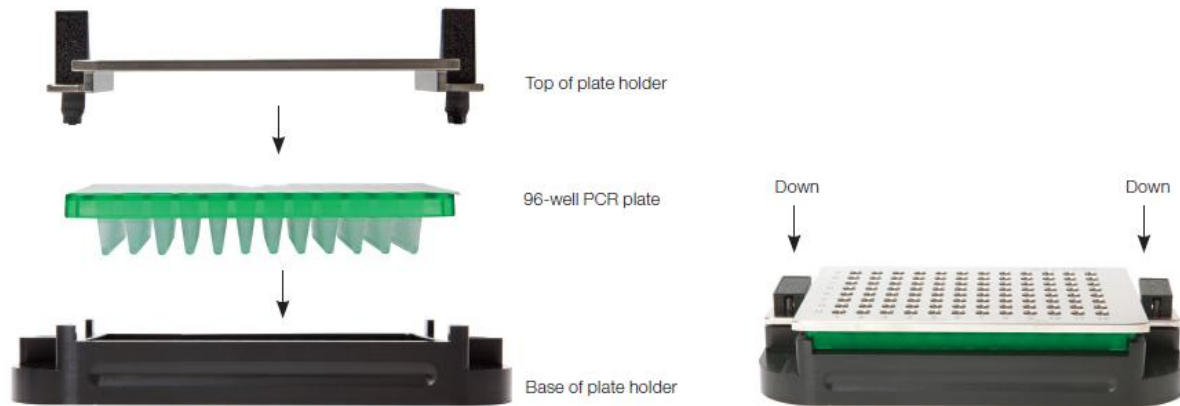


- ✓ Place holder on a flat surface
- ✓ Position pipet tips at a 30-45° angle
- ✓ Slowly draw 40ul of droplets into the pipet tips (~5 sec)

- ✓ Position the pipet tip along the side of the well and near the bottom of well
- ✓ Slowly dispense the droplets (~5 sec)

- ✓ 180°C 5sec

4. Place the holder into the reader



Placing the 96-well plate into the plate holder.



Placing the plate holder into the droplet reader.

5. Quantasoft software setup interface

File name

Load settings and data from previous experiments

Options for advanced data analysis and setup

Open experiment editor

Load or create a template (settings only)

Define experiment settings

Start the run

View and analyze data

Abort run or exit program

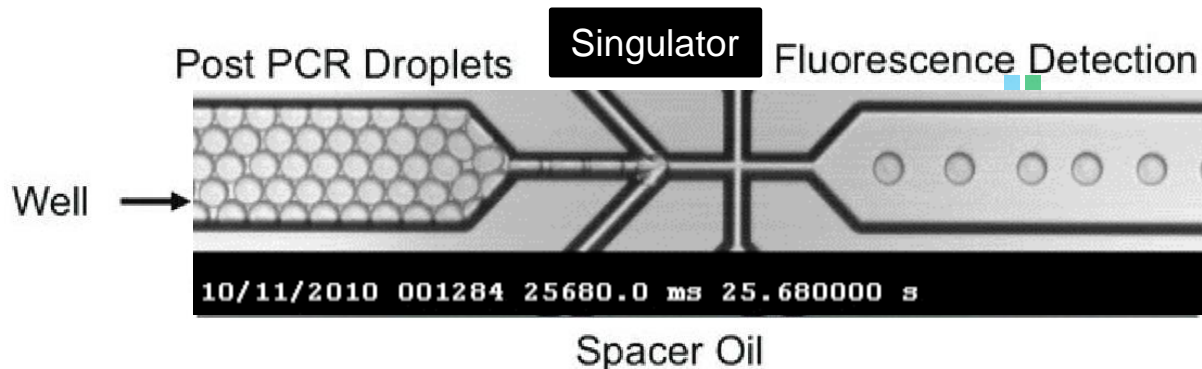
Instrument maintenance options (appear only when connected to the droplet reader or when in simulation mode)

Plate map

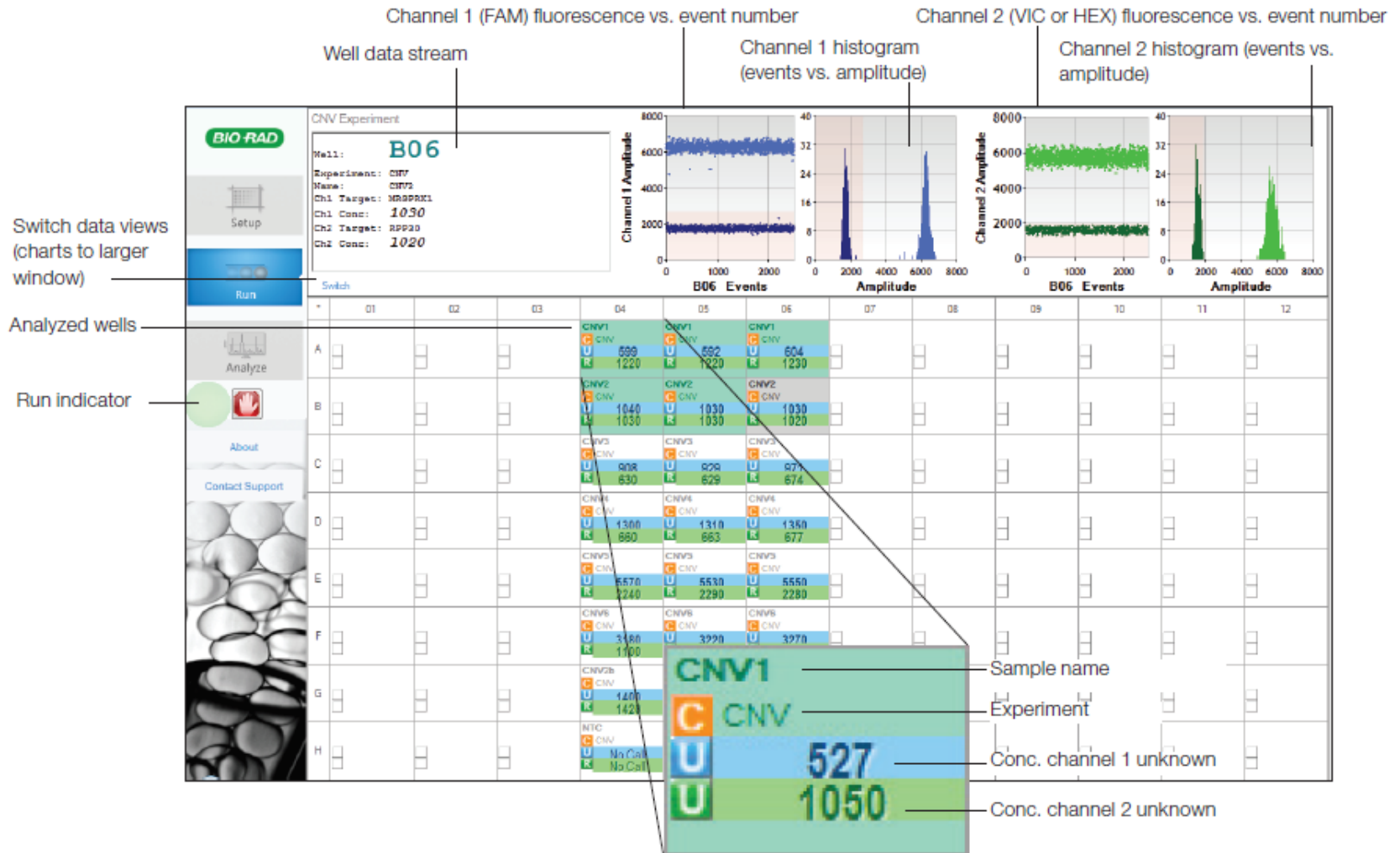
	01	02	03	04	05	06	07	08	09	10	11	12
WT	WT	WT	WT	CNV1	CNV1	CNV1	3180	3180	3180	4096	4096	4096
A	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	1049	1316	1069	545	533	550	3518	754	1579	3548	3838	3559
U	1049	1316	1069	510	519	519	1249	1249	1249	1249	1249	1249
Mutant (hom...)	Mutant (hom...)	Mutant (hom...)	CNV2	CNV2	CNV2	2948	2948	2948	1024	1024	1024	1024
B	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	908	337	1069	945	938	925	1179	1994	1999	994	994	998
U	8432	0	8432	925	949	932	1959	1959	1959	1239	1239	1239
90% mutant	90% mutant	90% mutant	CNV3	CNV3	CNV3	912	912	912	296	296	296	296
C	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	1969	2940	1969	876	844	883	487	482	473	237	237	232
U	1069	1920	1069	574	574	613	1289	1294	1289	1289	1270	1269
10% mutant	10% mutant	10% mutant	CNV4	CNV4	CNV4	108	108	108	64	64	64	64
D	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	1129	1120	1130	1990	1199	1199	197	120	196	96.5	99.2	96.7
U	1029	1930	1039	691	693	616	1359	1294	1294	1269	1240	1239
1% mutant	1% mutant	1% mutant	CNV5	CNV5	CNV5	32	32	32	16	16	16	16
E	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	1069	1940	1059	5870	5029	5950	38.5	26.9	36.7	14.7	13.6	14.4
U	1059	1930	1049	2840	2689	2660	1259	1279	1279	1239	1240	1239
0.1% mutant	0.1% mutant	0.1% mutant	CNV6	CNV6	CNV6	8	8	8	4	4	4	4
F	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	1099	1950	1049	2890	2539	2950	7.71	7.33	7.3	3.50	4.33	3.29
U	1099	1950	1049	396	1099	794	1259	1264	1279	1259	1240	1249
0.01% mutant	0.01% mutant	0.01% mutant	CNV7	CNV7	CNV7	2	2	2	1	1	1	1
G	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	962	399	964	4270	1289	1200	2.26	1.82	1.81	1.91	1.83	1.99
U	962	399	964	1290	1289	1200	1239	1244	1249	1249	1230	1239
NTC	NTC	NTC	NTC	NTC	NTC	NTC	0.5	0.5	0.5	0	0	0
H	RED	RED	RED	CNV	CNV	CNV	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series	AGG Di Series
U	9	0	0	No Ctrl	No Ctrl	No Ctrl	9392	6492	8695	9	9	9
U	9	0	0	No Ctrl	No Ctrl	No Ctrl	1229	1234	1239	1249	1220	1269

Droplet Reading

- Autosampler of QX200 Droplet Reader processes each sample independently
- Droplets stream single-file past the optical detector (32 wells/hr in 2 color)

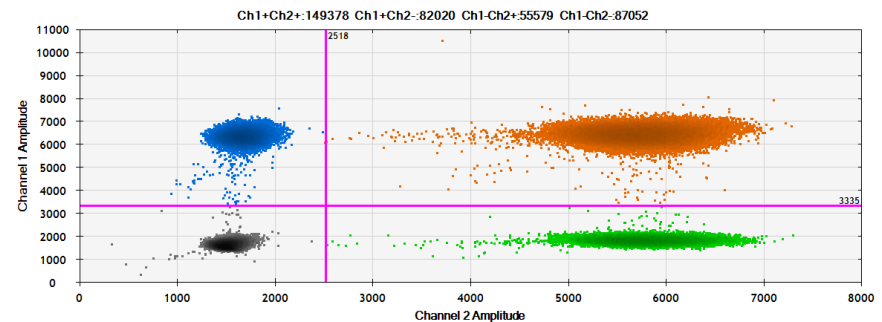
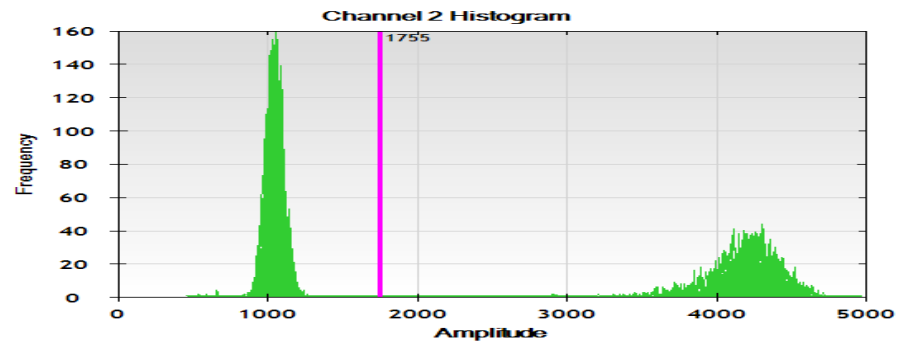
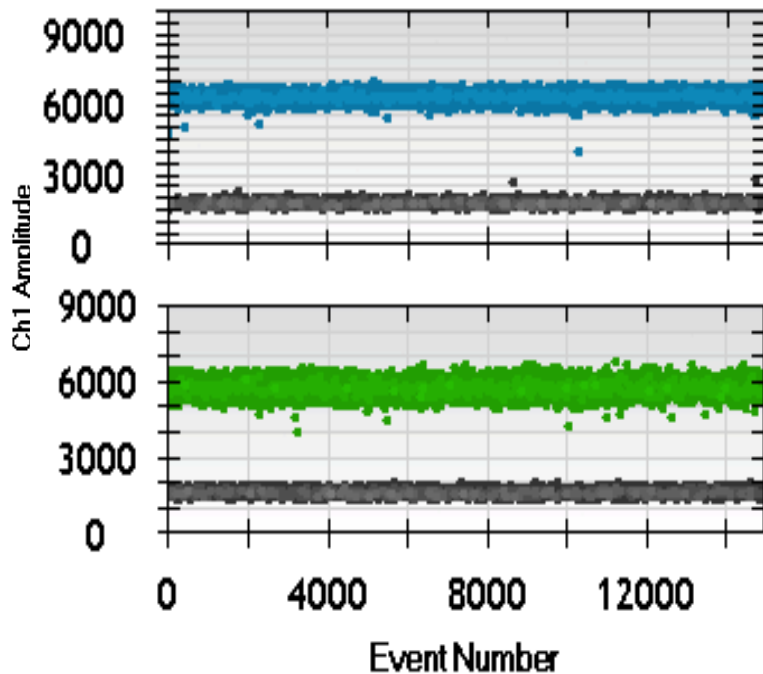


Quantasoft software interface

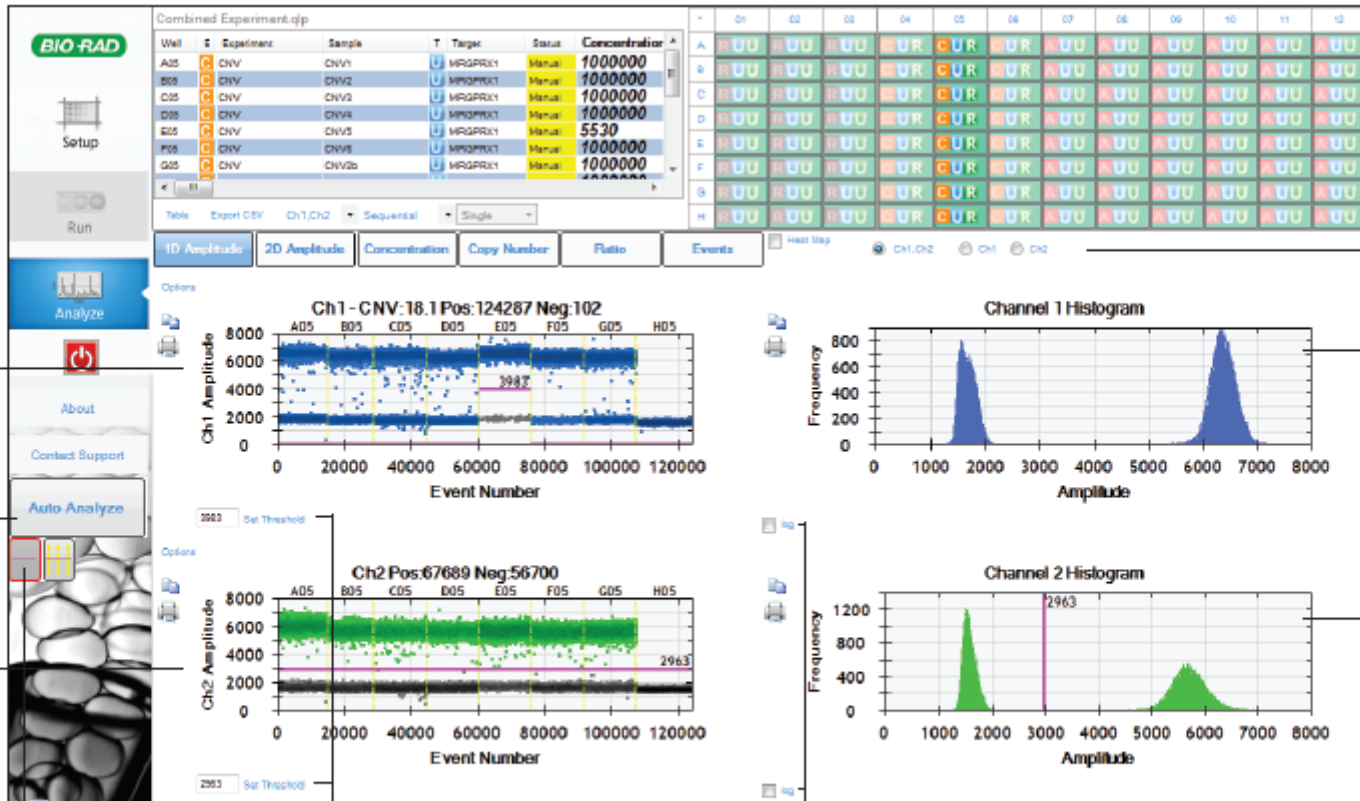


Droplet Fluorescence Converted to a Digital Signal

- Positive droplets contain at least 1 copy of target DNA (cDNA)
- Positive droplets have increased fluorescence vs. negative droplets
- QuantaSoft™ software measures the number of positive and negative droplets per fluorophore per sample



6. Analysis



Channel 1 fluorescence amplitude vs. event number

Reset automatic threshold settings

Channel 2 fluorescence amplitude vs. event number

Threshold adjustment tools

Threshold settings

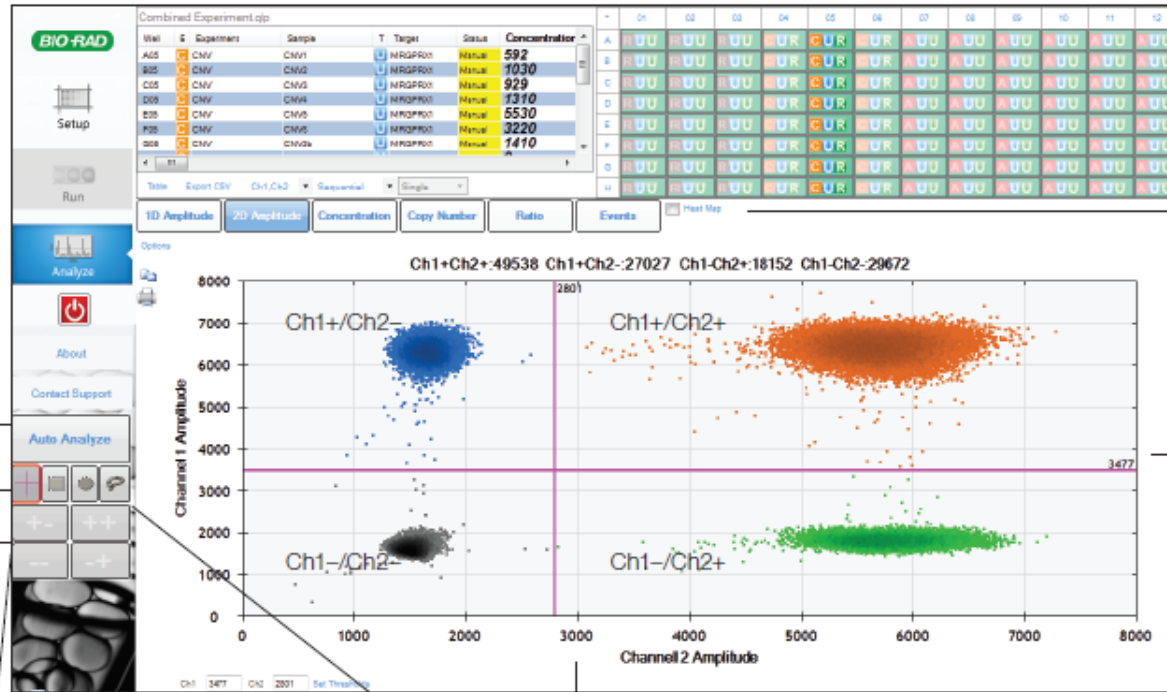
Y-axis log scale toggle

Channel selector

Channel 1 (FAM) histogram of events vs. amplitude

Channel 2 (VIC or HEX) histogram of events vs. amplitude

6. Analysis



Display multicolor heat map

Reset automatic threshold settings
 Threshold adjustment tools
 Working cluster selector

Ch 1 threshold

Threshold settings

Ch2 threshold

Ch1+/Ch2-



Ch1+/Ch2+

Ch1-/Ch2-



Ch1-/Ch2+

6. View Concentration Data



Select channel(s) to display

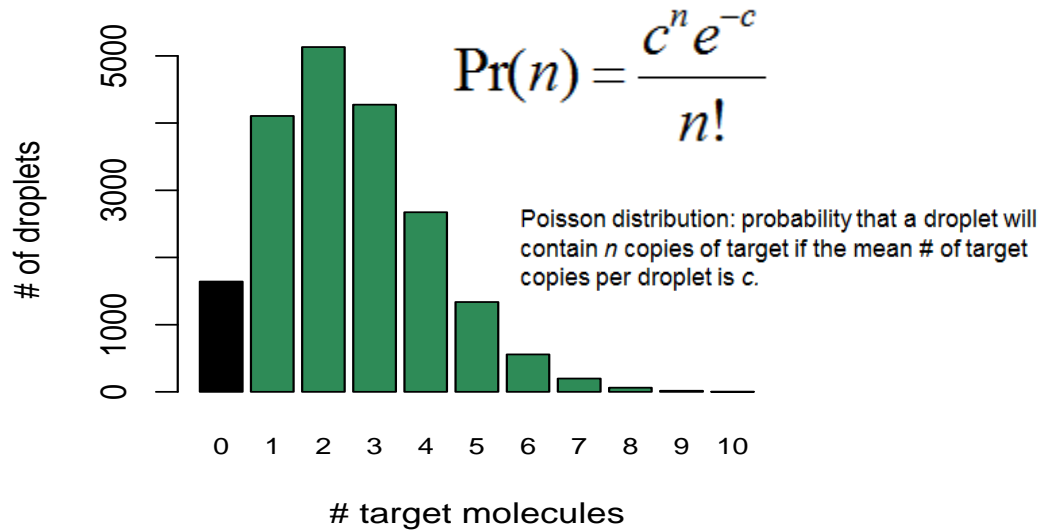
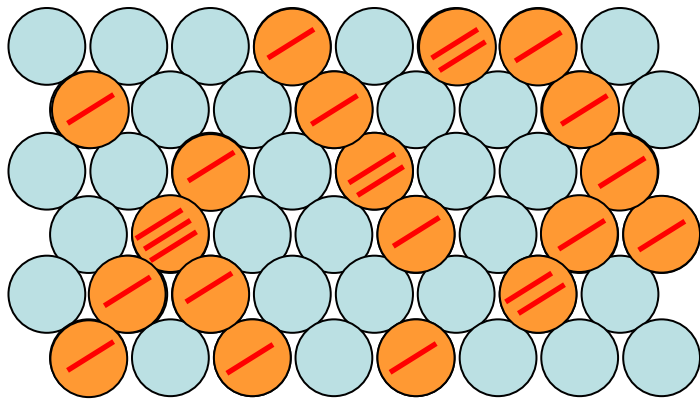
- Y2 Axis options:
- None** — no Y2 axis
 - Ch2** — display Ch2 concentration
 - CNV** — display copy number (CNV analysis only)
 - Ratio(a/b)** — ratio (a/b) of unknown:reference
 - Fractional Abundance (a/(a+b))** — ratio (a/[a+b]) of unknown:reference; abundance
 - Inverse** — inverse of ratio or fractional abundance

Toggle y-axis log scale

Display data from single wells, merged data, or both

Toggle x-axis values and well, name, or label display

Beyond Limiting Dilution: High Target Concentration (with ddPCR)



Siméon Denis Poisson
(1781-1840)

“X” target
copies

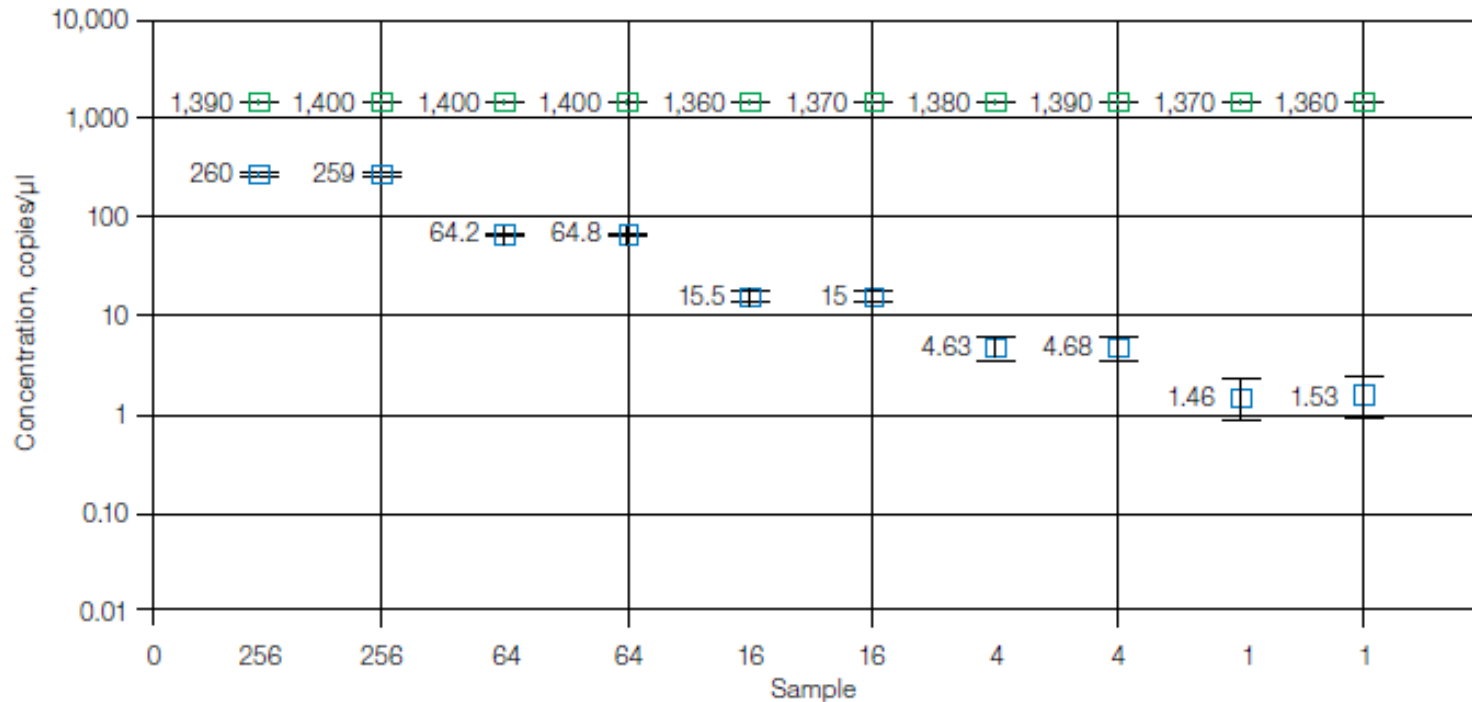
Modeling as Poisson

Copies per droplet = $-\ln(1 - p)$

where p = fraction of positive droplets **and**
 $(1 - p)$ = fraction of negative droplets.

Note: At 1 target copy/droplet, 37% negatives!

Excellent Reproducibility and Linearity Across Concentrations and Instruments



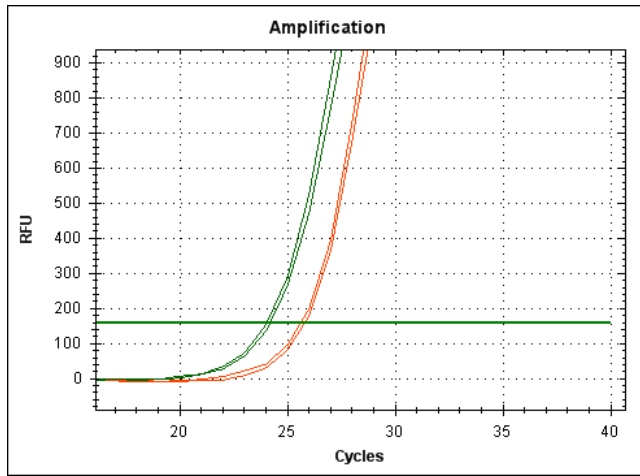
- Readout: copies/μl
- Dynamic range: 1–100,000 copies/well (~330 ng human genomic DNA)



Benefits of Droplet Digital PCR

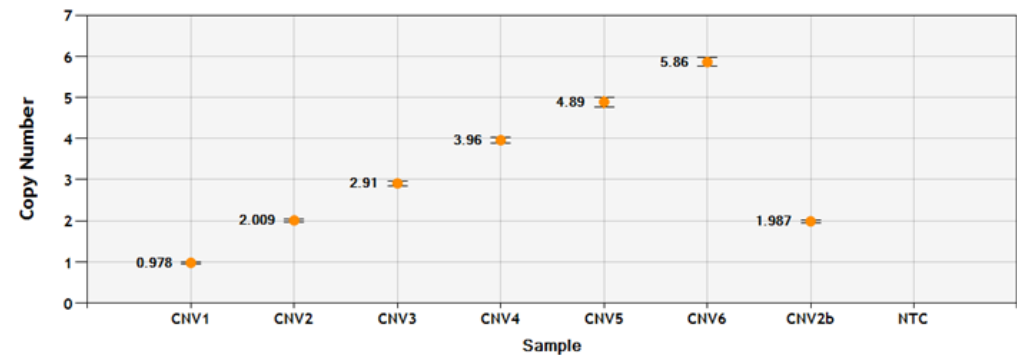
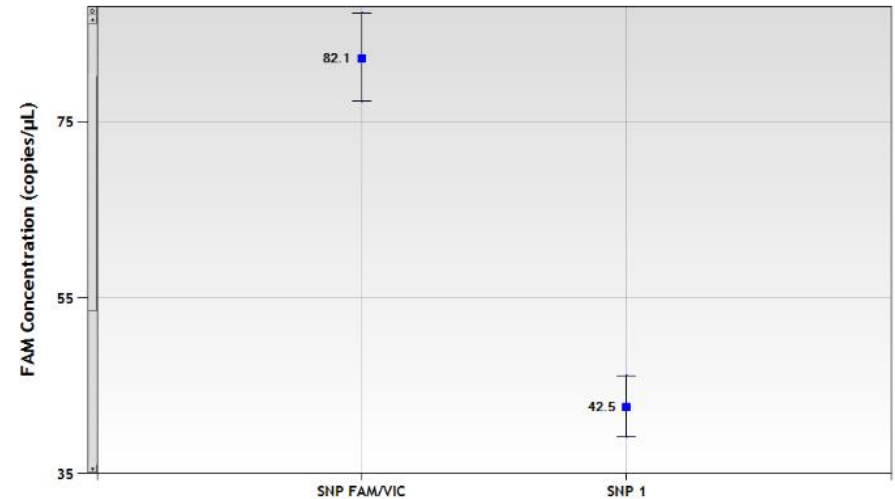
Detecting Small Fold Changes

A 2-fold change measured by real-time PCR



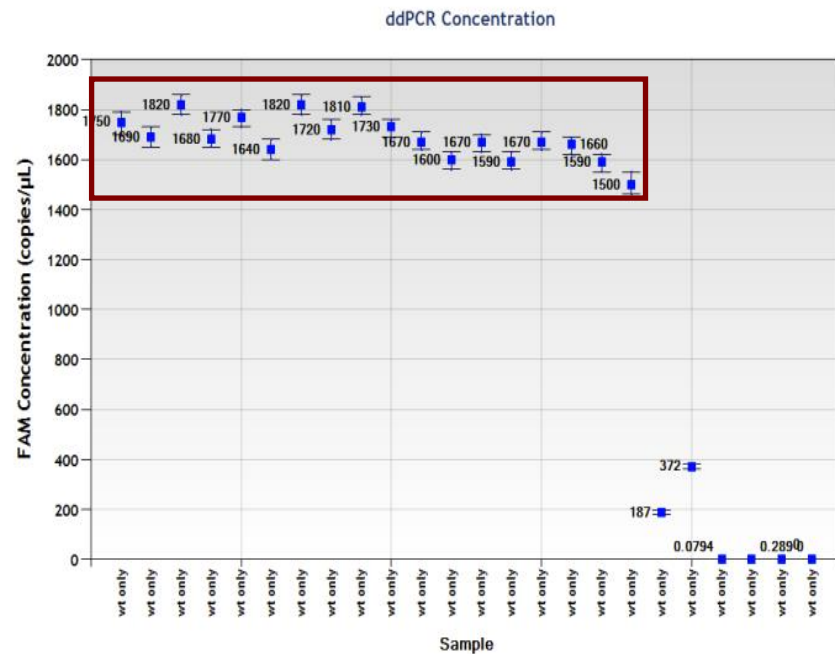
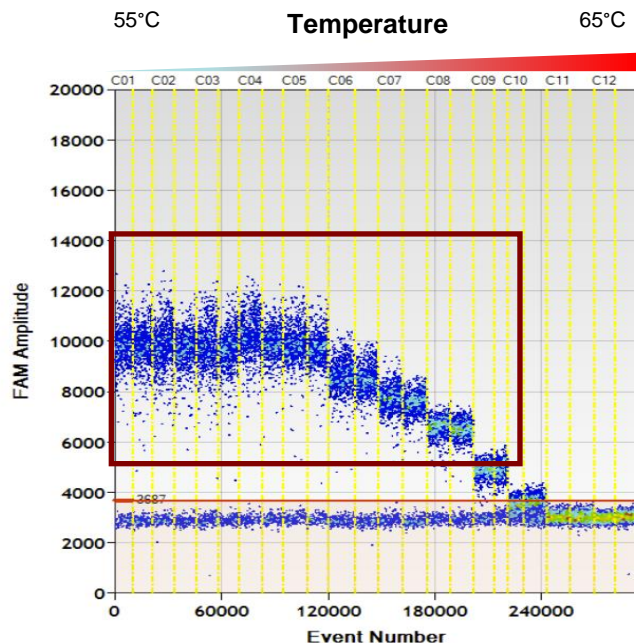
Content	Cq	Cq Std. Dev
Unkn-14	26.99	0.110
Unkn-14	26.84	0.110
Unkn-15	25.47	0.115
Unkn-15	25.31	0.115

A 2-fold change measured by ddPCR



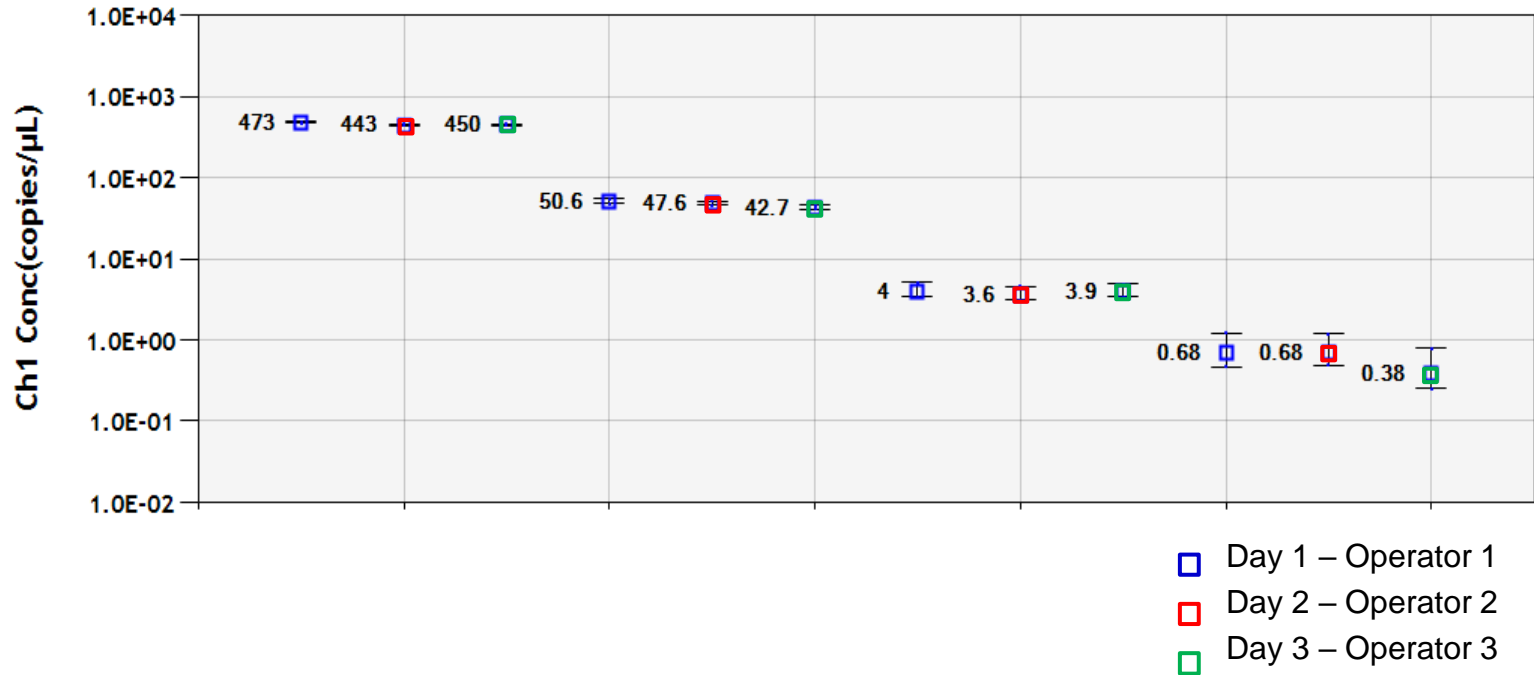
PCR efficiency has a minimal impact on ddPCR[†]

- Thermal gradient mimics different PCR efficiencies
- Same concentration measured across different annealing temps



ddPCR[†] reproducibility

- ✓ Reproducibility trial – 3 operators in 3 different Korean laboratories using the same assay and samples obtained a high degree of data uniformity





Applications of Droplet Digital PCR

Applications of ddPCR



▪ Cancer Biomarker Studies and Copy Number Variation

- Measure varying degrees of cancer mutations, detect rare DNA target copies, and resolve copy number variation states with superior sensitivity and resolution.



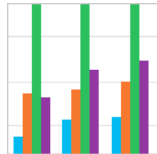
▪ Pathogen Detection

- Employ the extreme precision of the QX200 System to quantify small fold changes in target DNA or RNA molecules in pathogen detection and monitoring.



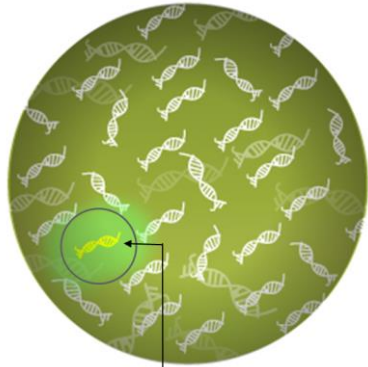
▪ Next Generation Sequencing

- Perform accurate quantification and qualification of NGS libraries
- NGS data validation



▪ Gene Expression Analysis

- Achieve reliable and reproducible measurements of small fold changes for low abundance of mRNA and microRNA and RNA variants



Target DNA



▪ Food Testing

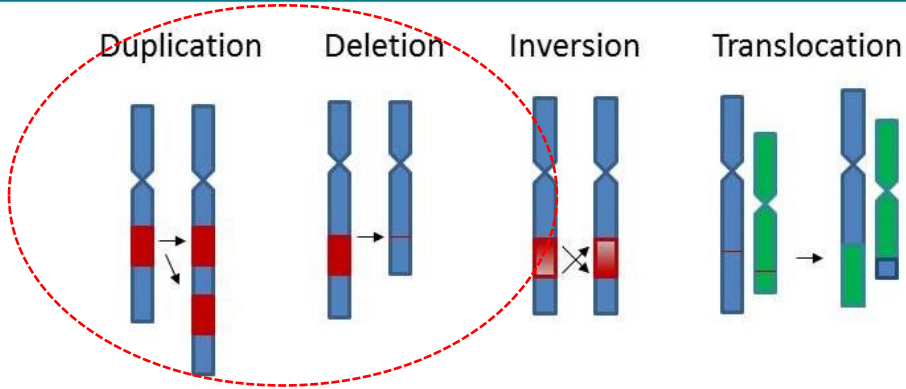
- Perform routine evaluation of genetically modified organisms (GMO) using validated ddPCR methods.



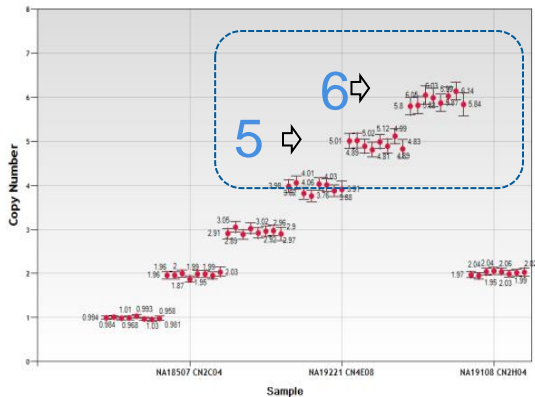
▪ Environmental Monitoring

- Test a wide variety of environmental samples like soil and water

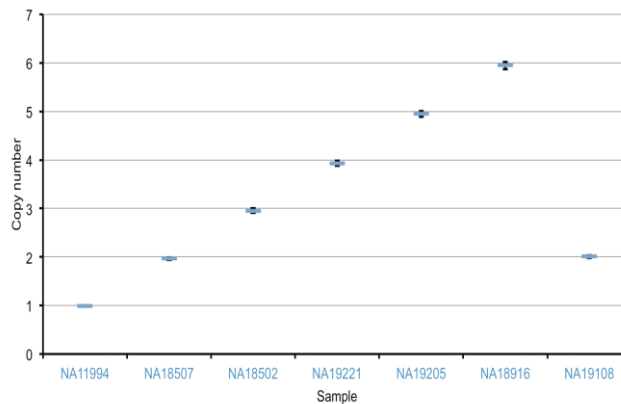
Copy Number Variation (CNV's)



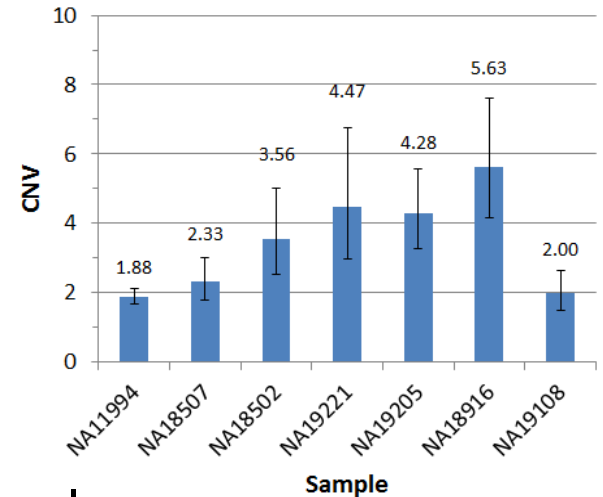
ddPCR individual wells



ddPCR merged wells



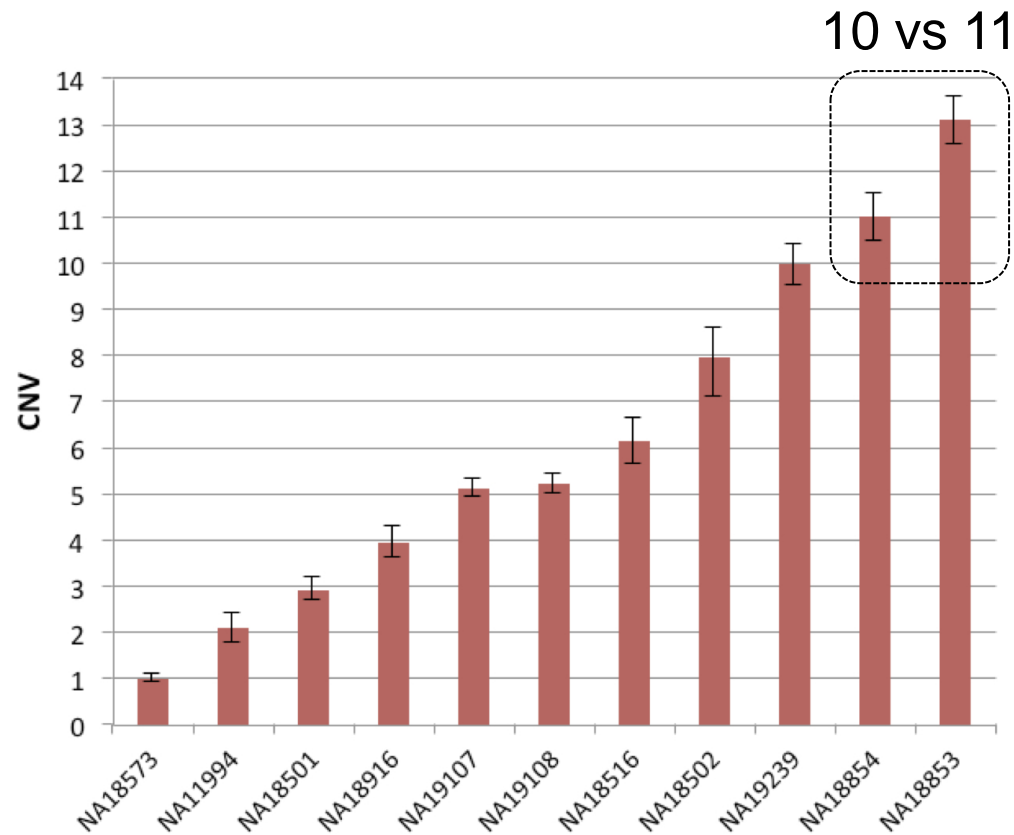
real-time PCR (8 wells)



- Measure *MRGPRX1* gene copies from various samples

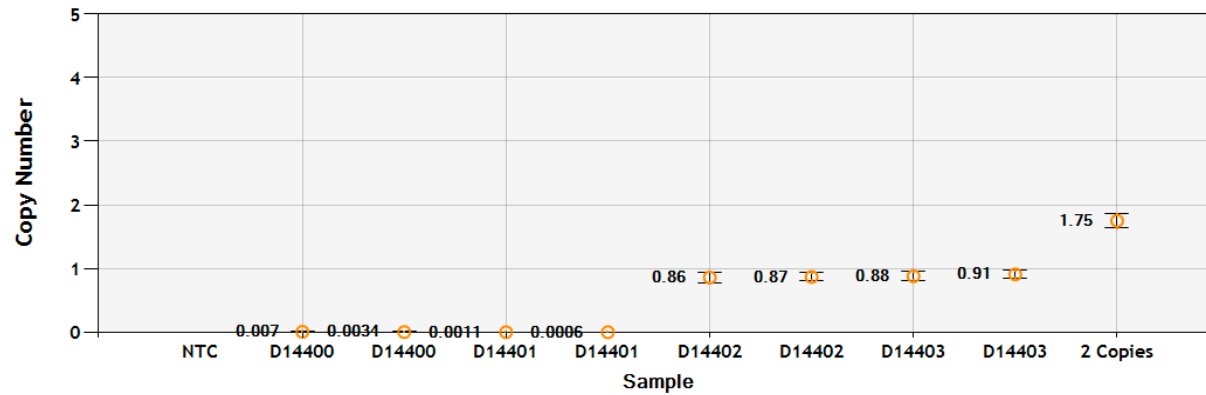
Higher CNV Level Discrimination

Resolution of **10 vs. 11 copies** of CCL3L1 from HapMap samples

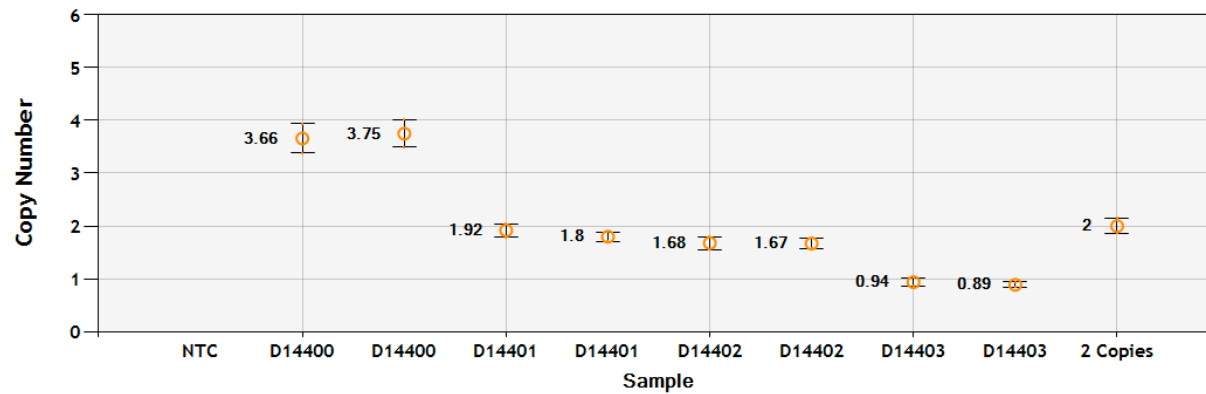


SMN1, SMN2 Copy Number Determinant Kit

SMN1



SMN2

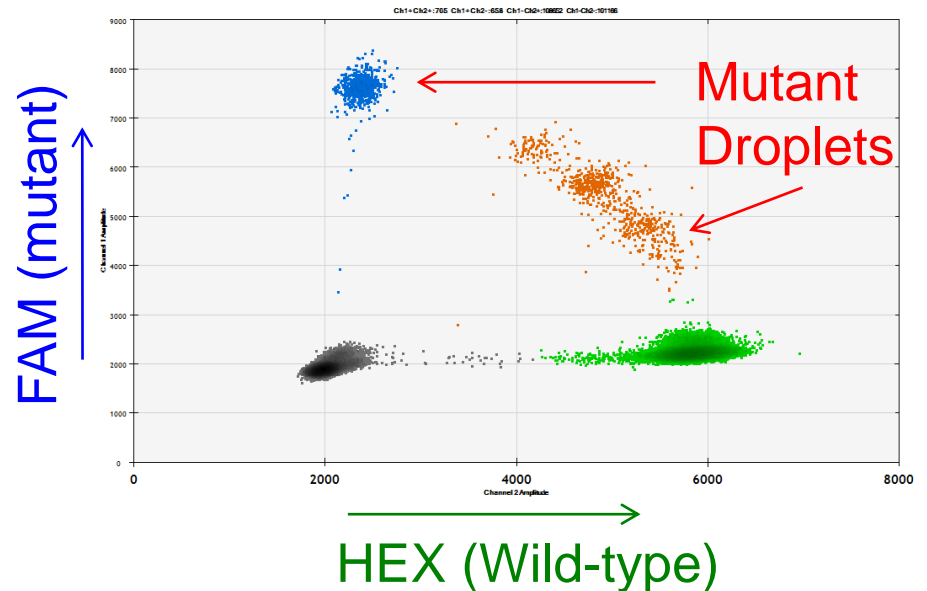
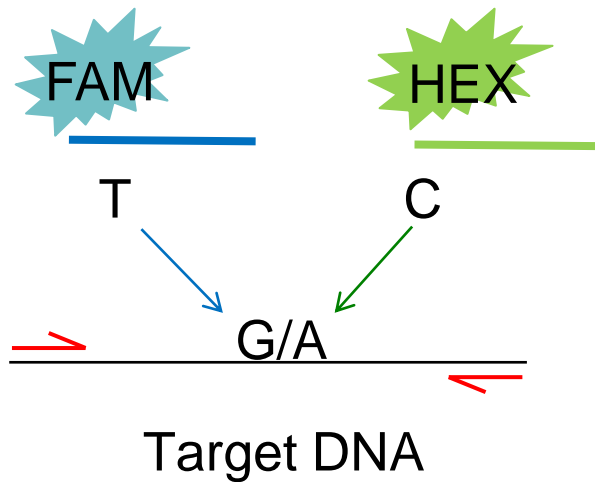


Rare Event Detection Assays

Rare mutation detection assays share common primers while the **probes** are **SNP-dependent**

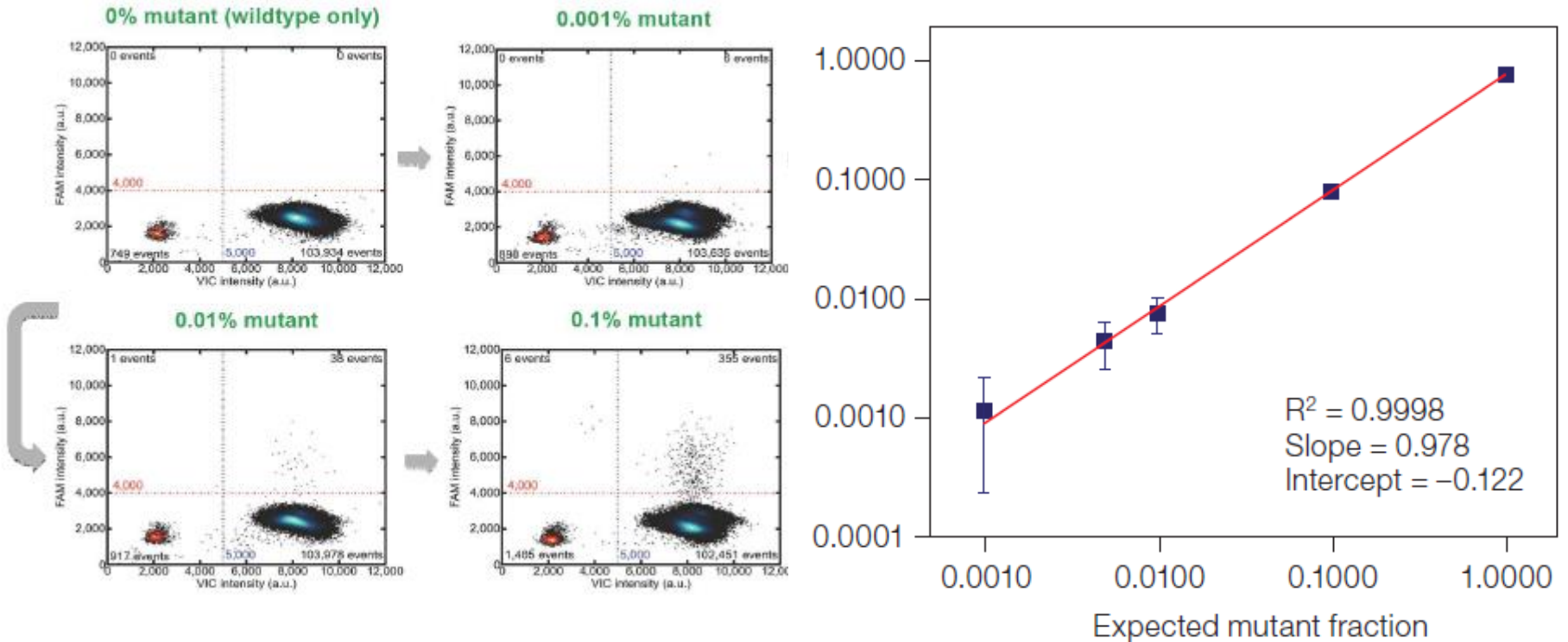
Mutant Probe

Wild-type Probe



Rare Event Detection

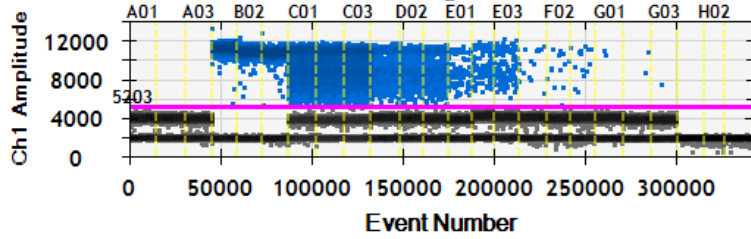
BRAF V600E ddPCR Detected Down to 1/100,000



qPCR detects *BRAF V600E* down to 2.0%

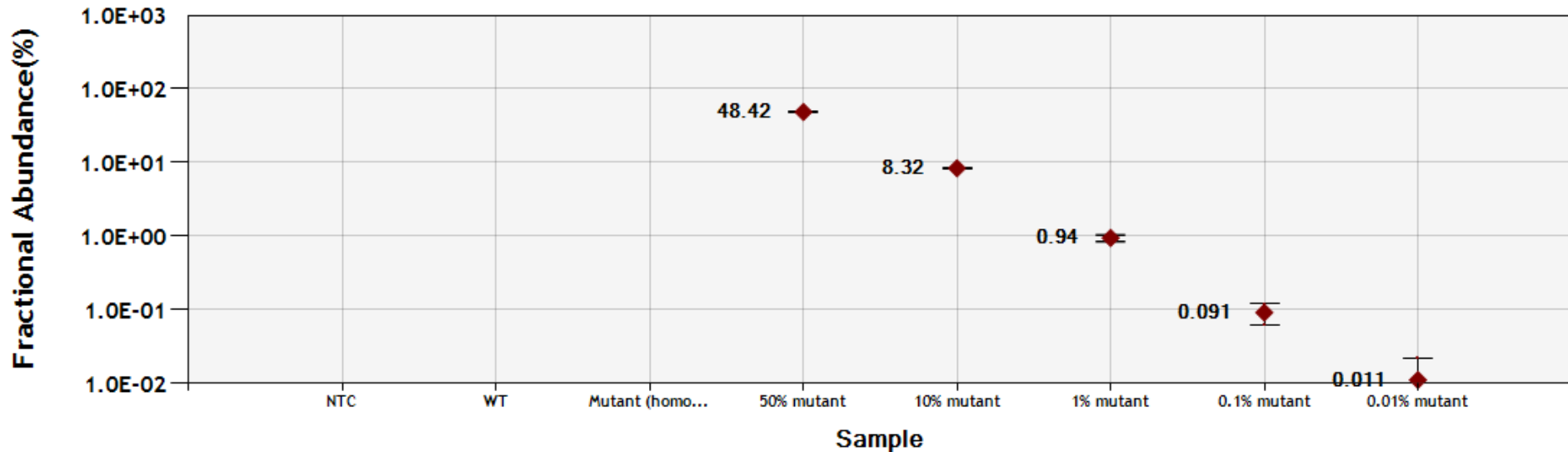
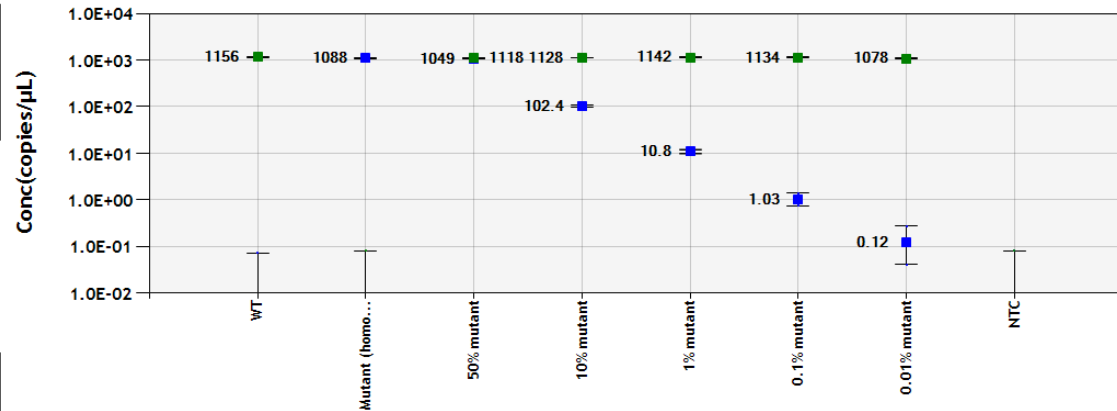
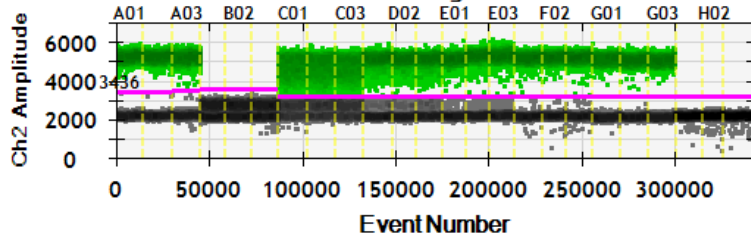
BRAF V600E detection by ddPCR sensitivity 0.01%

Ch1 Pos:58327 Neg:283609



5203 Set Threshold

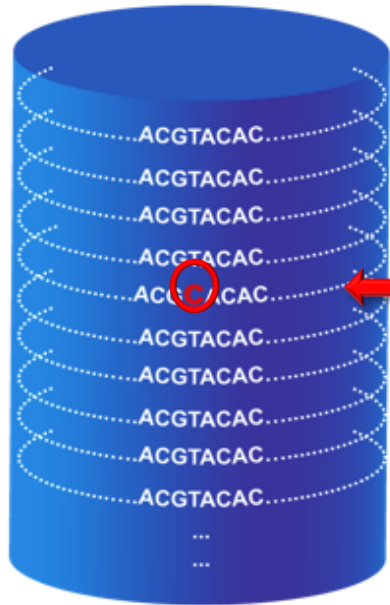
Ch2 Pos:165896 Neg:176040



Droplet Partitioning Increases Mutant Abundance

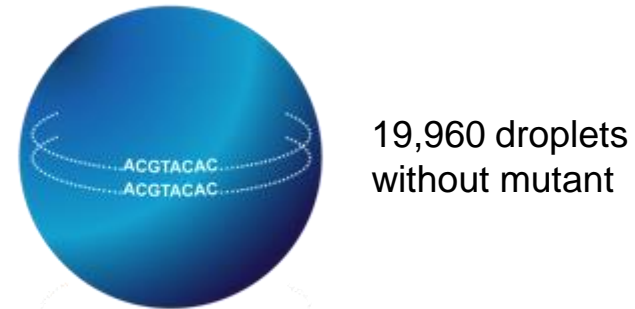
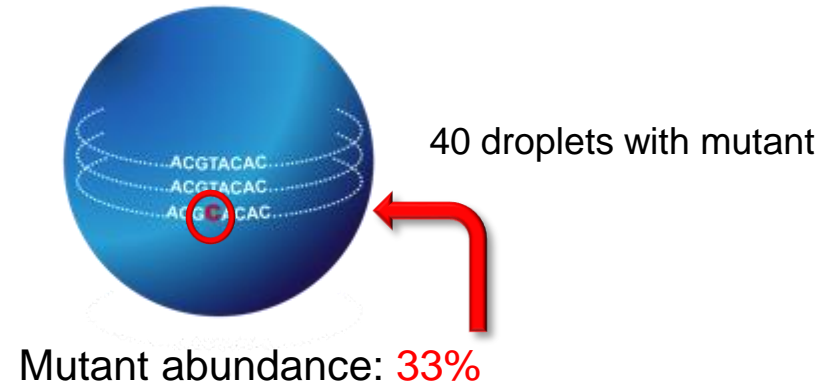
qPCR Reaction — 1 x 20 μ l

40,000 wild-type molecules
40 mutant molecules

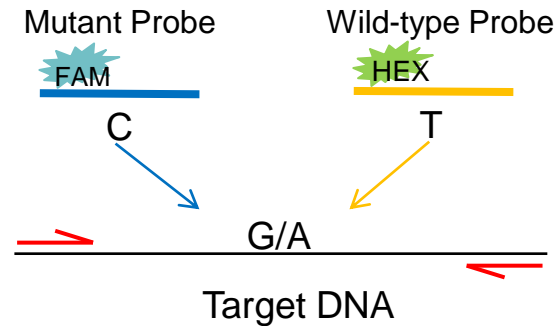


Mutant abundance: 0.1%

ddPCR Partitioned Reaction — 20,000 x 1 nl

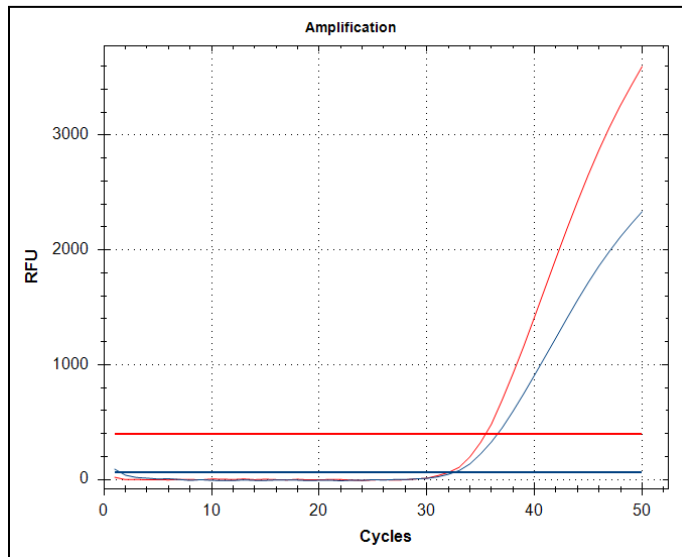


ddPCR assays are more easy to design and validate

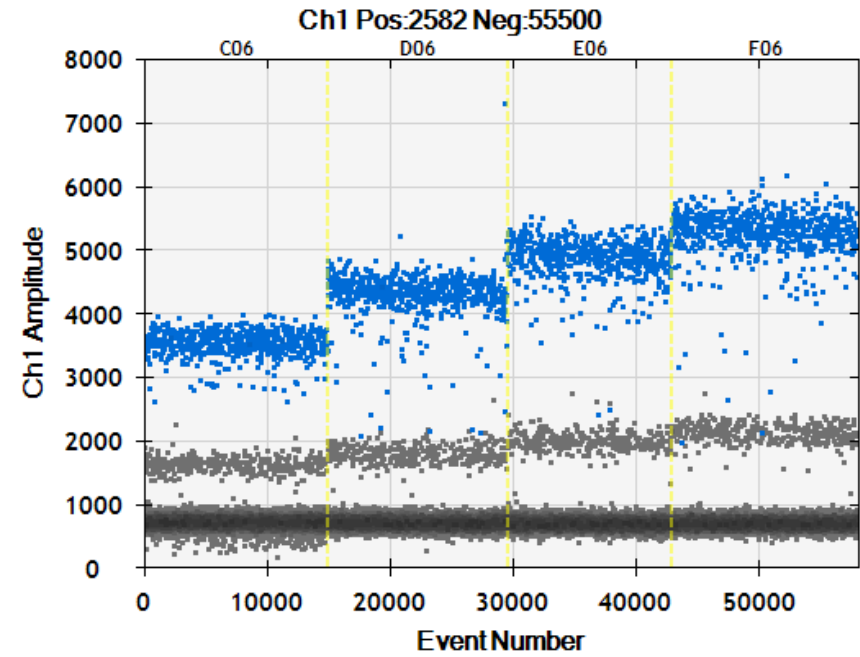


- More Tolerance to:
1. Non-specific binding
 2. Primer dimer
 3. PCR inhibitor

Non-specific cross reaction

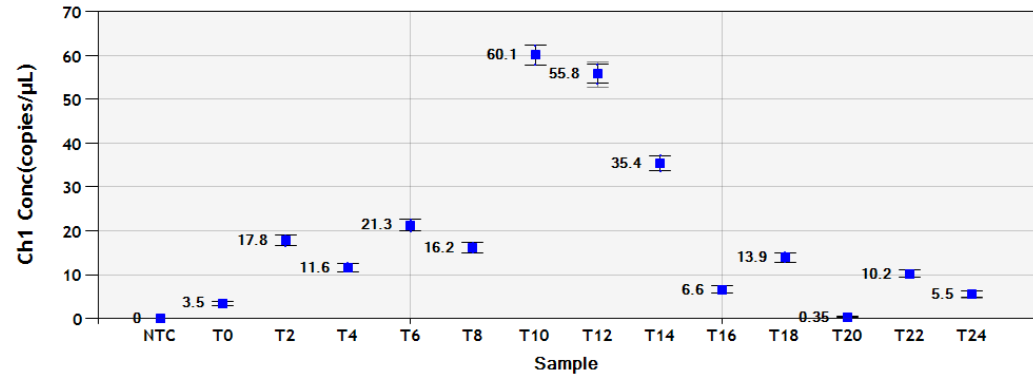
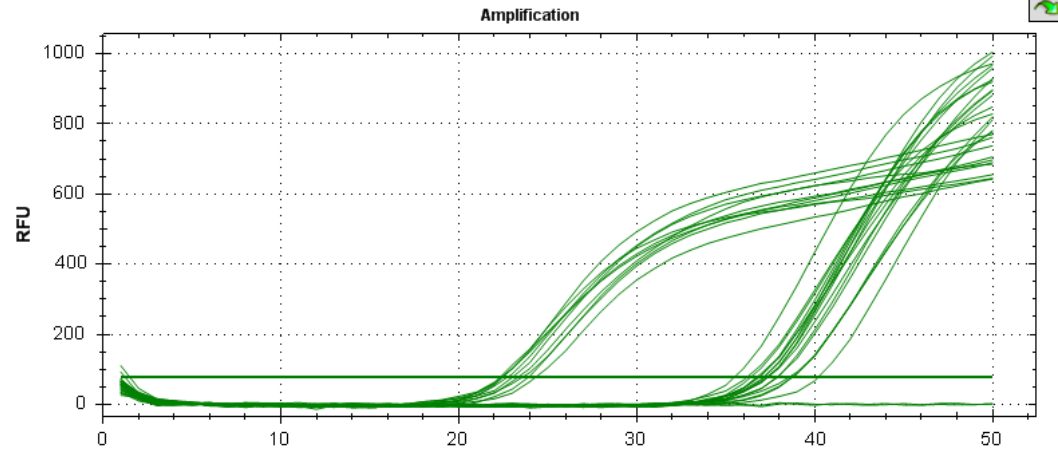
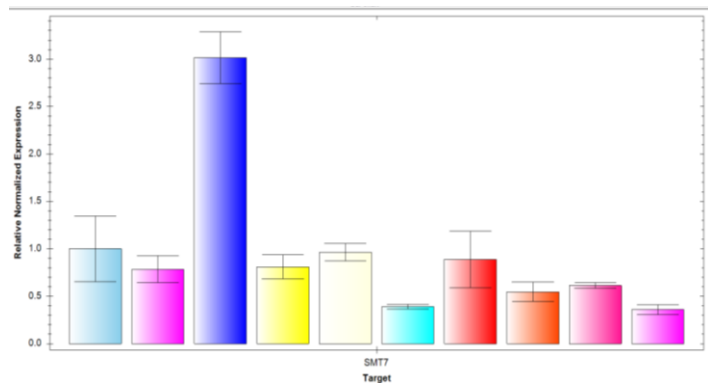
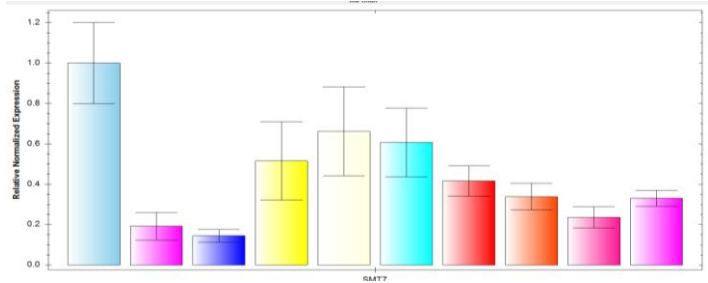
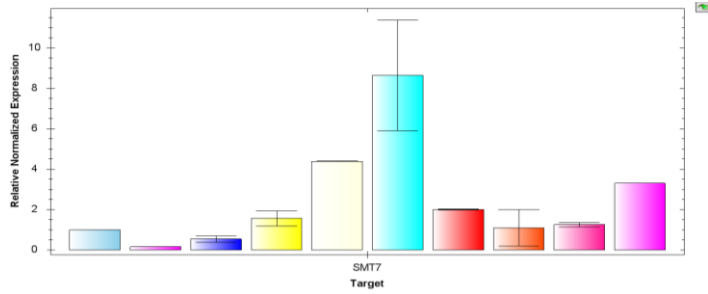


qPCR



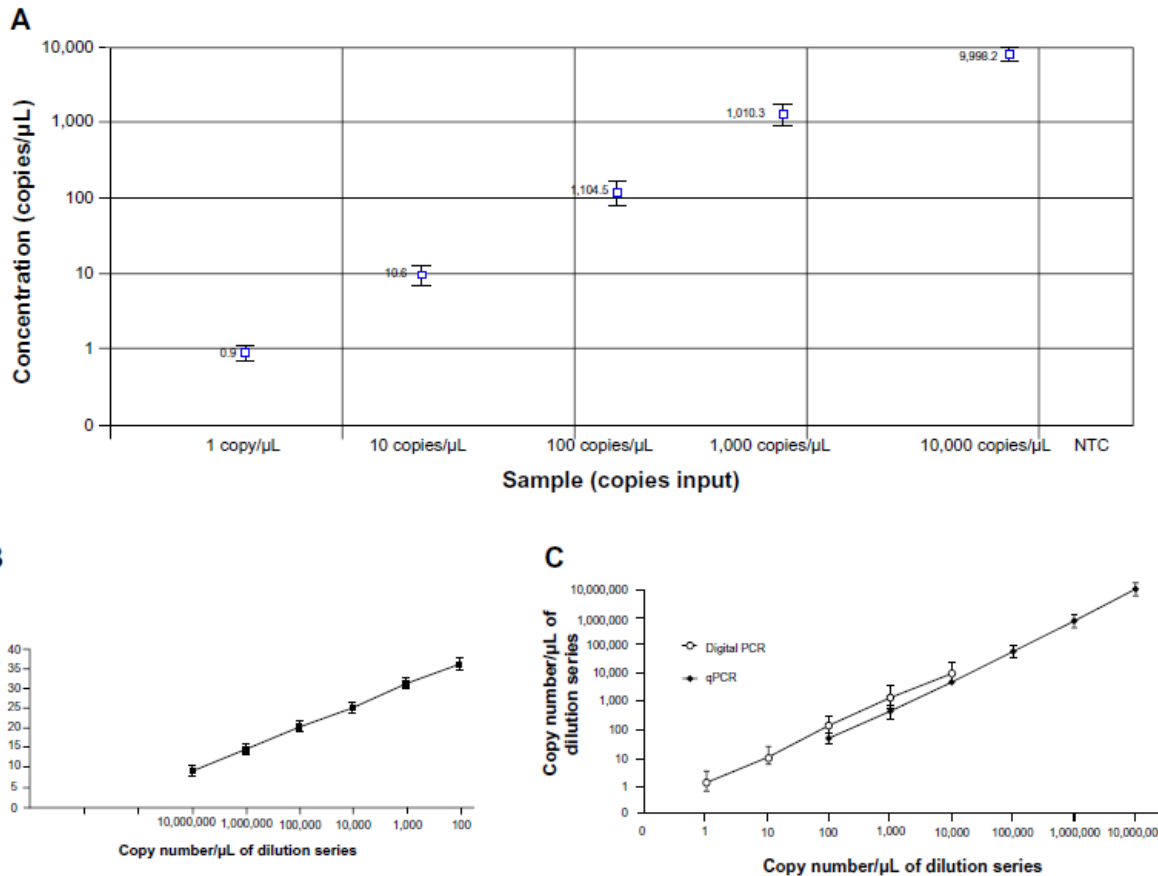
ddPCR

Gene Expression Q-PCR VS ddPCR



Gene expression

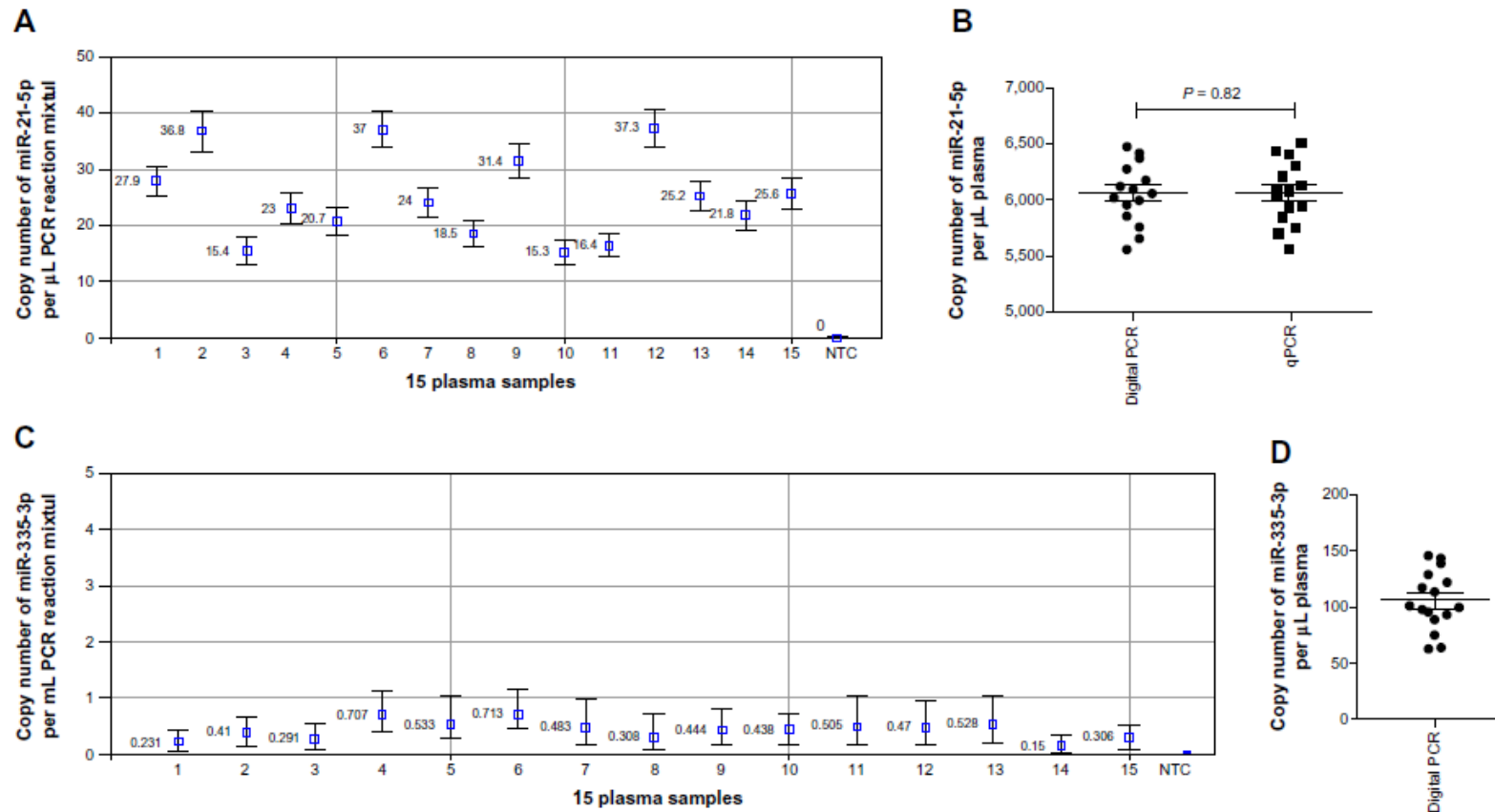
Quantification of Plasma miRNAs by Digital PCR for Cancer Diagnosis



Overall, digital PCR had a narrower dynamic range.

However, the lowest copy number detected by digital PCR was significantly lower (1 copy per μL of input) than that by qPCR (approximately 100 copies per μL of input)..

Quantification of Plasma miRNAs by Digital PCR for Cancer Diagnosis

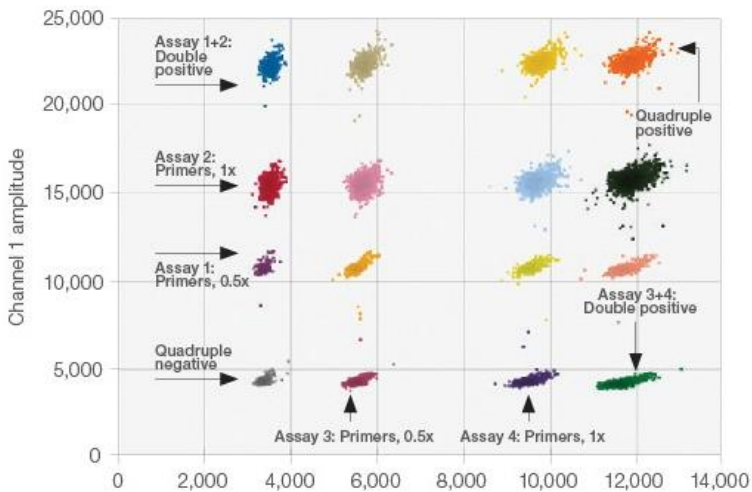


Therefore, digital PCR rather than qPCR might reliably and sensitively measure the copy number of miR-335-3p that has endogenous low-level expression in plasma.

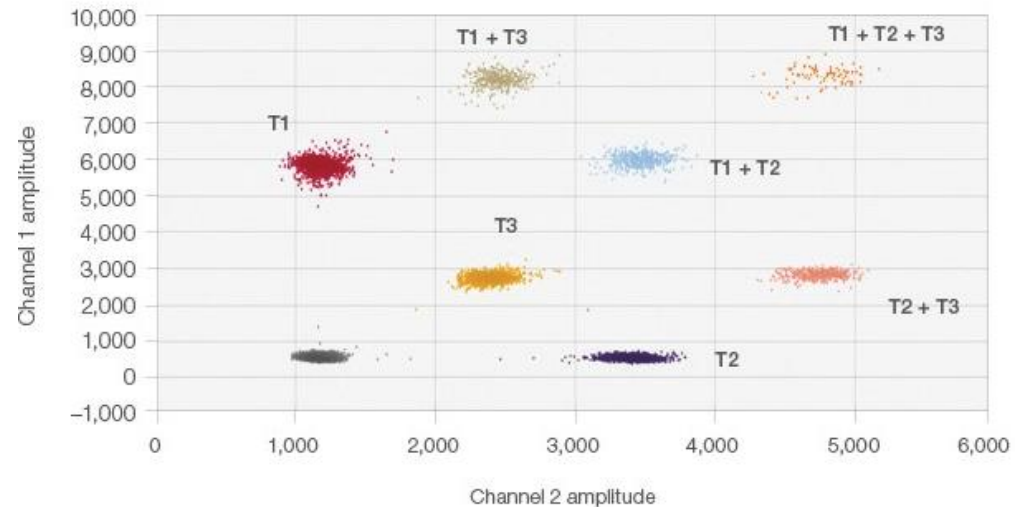
Expanded Droplet Digital PCR Multiplexing Capability

47

- Amplitude Multiplex Droplet Digital PCR

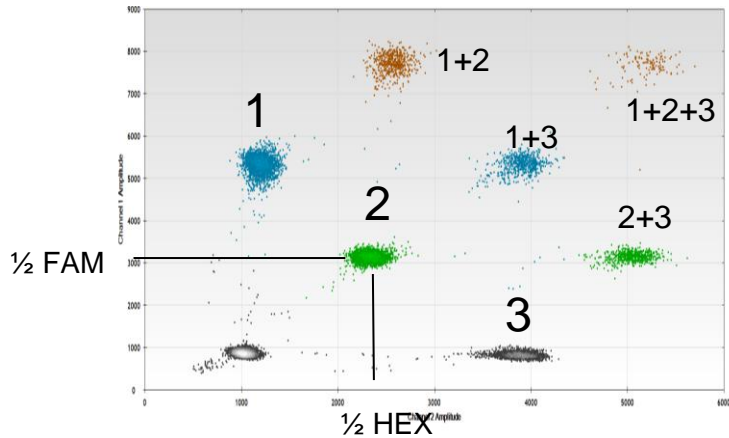


- Probe-Mixing Multiplexing Droplet Digital PCR



Multiplexing in 2 channels overview

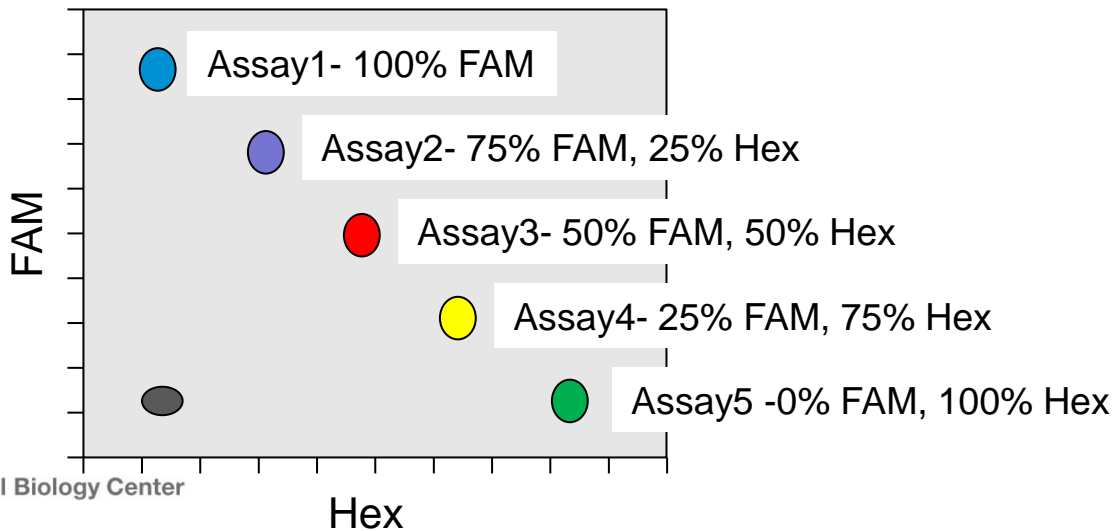
Concept of probe mixing - triplex



Double,
Triple
positives

3plex	Assay mixing
1	100% FAM, 0% Hex
2	50% FAM, 50% Hex
3	0% FAM, 100% Hex

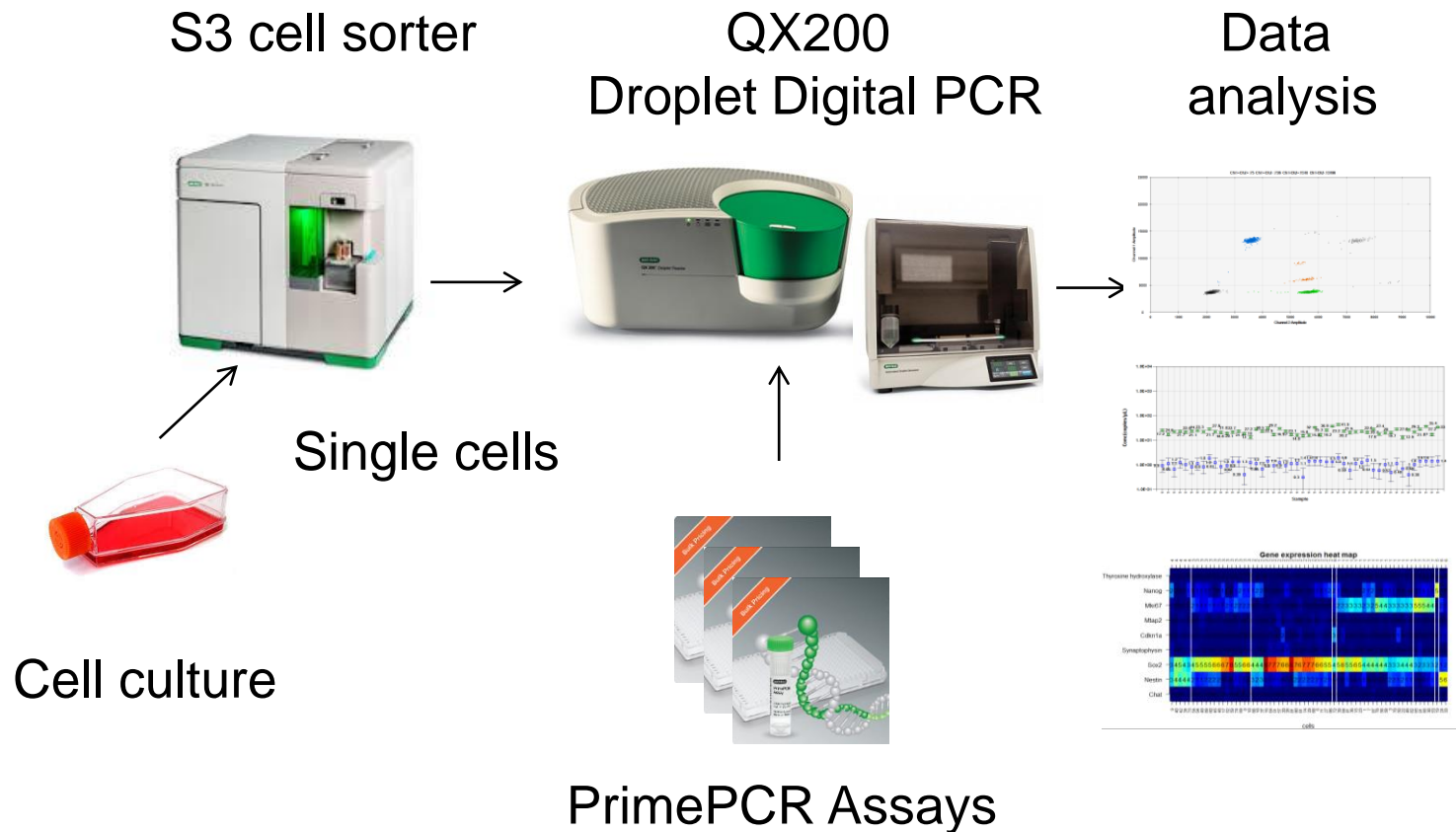
Theoretical clustering -5plex



5plex	Assay mixing
1	100% FAM, 0% Hex
2	75% FAM, 25% Hex
3	50% FAM, 50% Hex
4	25% FAM, 75% Hex
5	0% FAM, 100% Hex

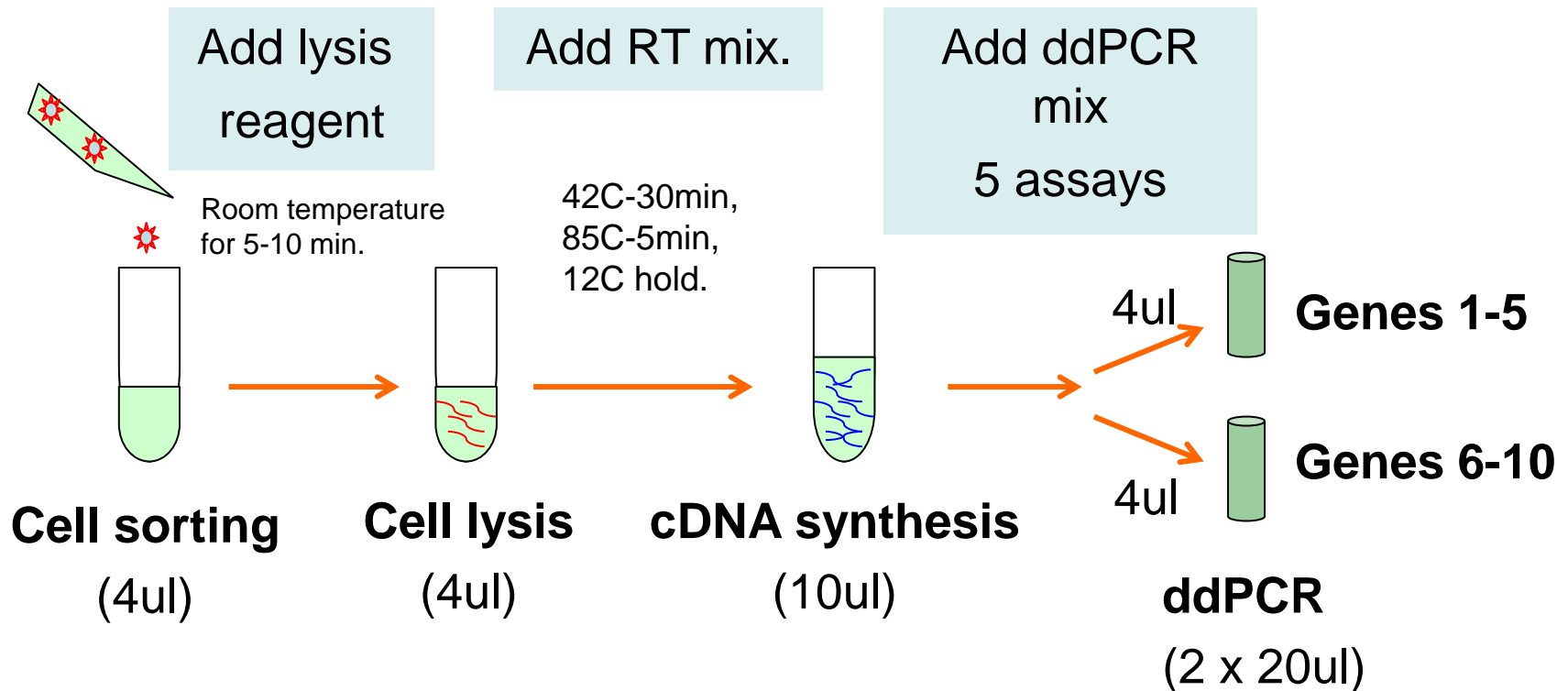
Single Cell Analysis Using ddPCR

Simplified workflow completed in less than 1 workday

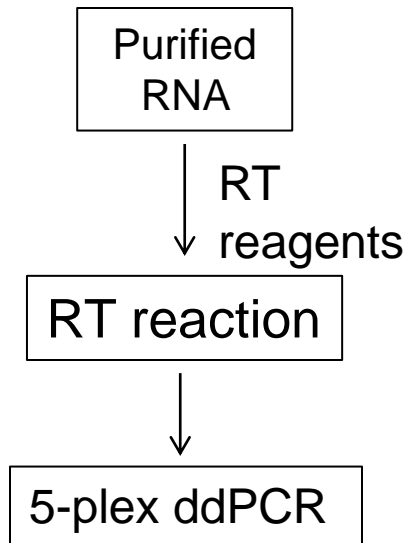


Single Cell Analysis Using ddPCR

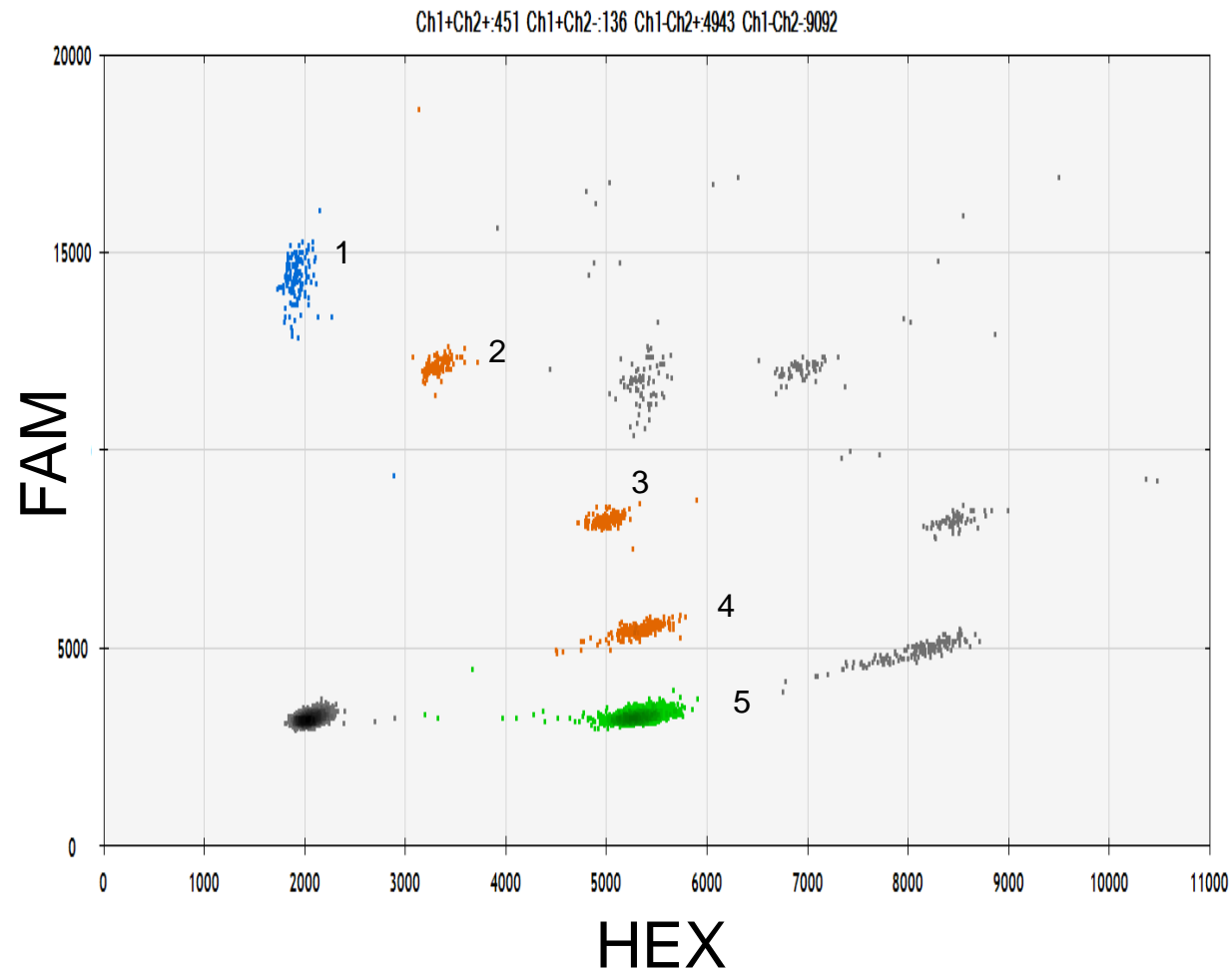
Measure 10 genes per single cell with **no pre-amp**



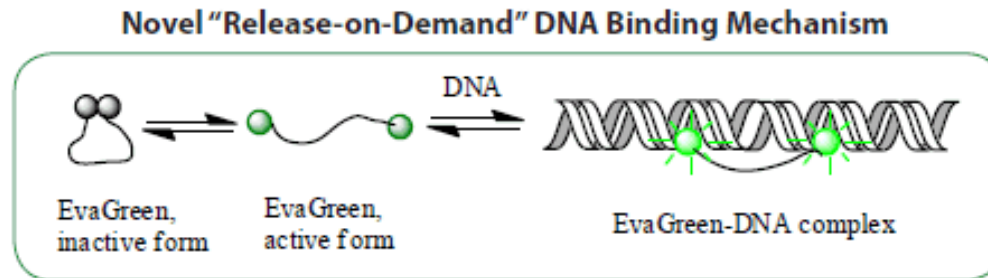
5-plexing with probe mixing



5plex	Gene	Assay mixing
1	Hoxa1	100% FAM
2	Oct4	75% FAM, 25% Hex
3	Nestin	50% FAM, 50% Hex
4	Sox2	25% FAM, 75% Hex
5	RPLP0	100% Hex



QX200 System Enables dsDNA Detection Capability with EvaGreen (without TaqMan probes)



- No preference for GC- or AT-rich sequence
- Less PCR inhibition than SYBR[®] Green and lower tendency to cause nonspecific amplification
- Tolerated at a higher concentration, which enables a brighter signal
- Good stability
- Safety
 - Dye is impenetrable to both latex gloves and cell membranes
 - Dye is noncytotoxic and nonmutagenic at concentrations used in the laboratory

High Sensitivity Detection and Quantitation of DNA Copy Number and Single Nucleotide Variants with Single Color Droplet Digital PCR

High Sensitivity Detection and Quantitation of DNA Copy Number and Single Nucleotide Variants with Single Color Droplet Digital PCR

Laura Miotke,^{‡,†} Billy T. Lau,^{‡,§,†} Rowza T. Rumma,^{‡,†} and Hanlee P. Ji^{*,‡,§}

[‡]Division of Oncology, Department of Medicine, Stanford University School of Medicine, CCSR 1115, 269 Campus Drive, Stanford, California, 94305 United States

[§]Stanford Genome Technology Center, Stanford University, Palo Alto, California, 94304 United States

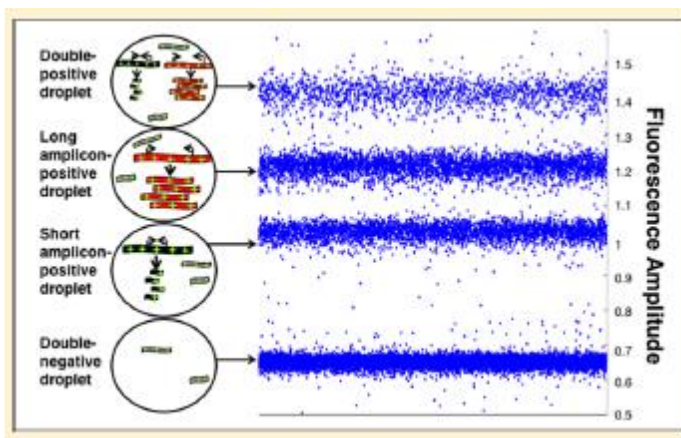
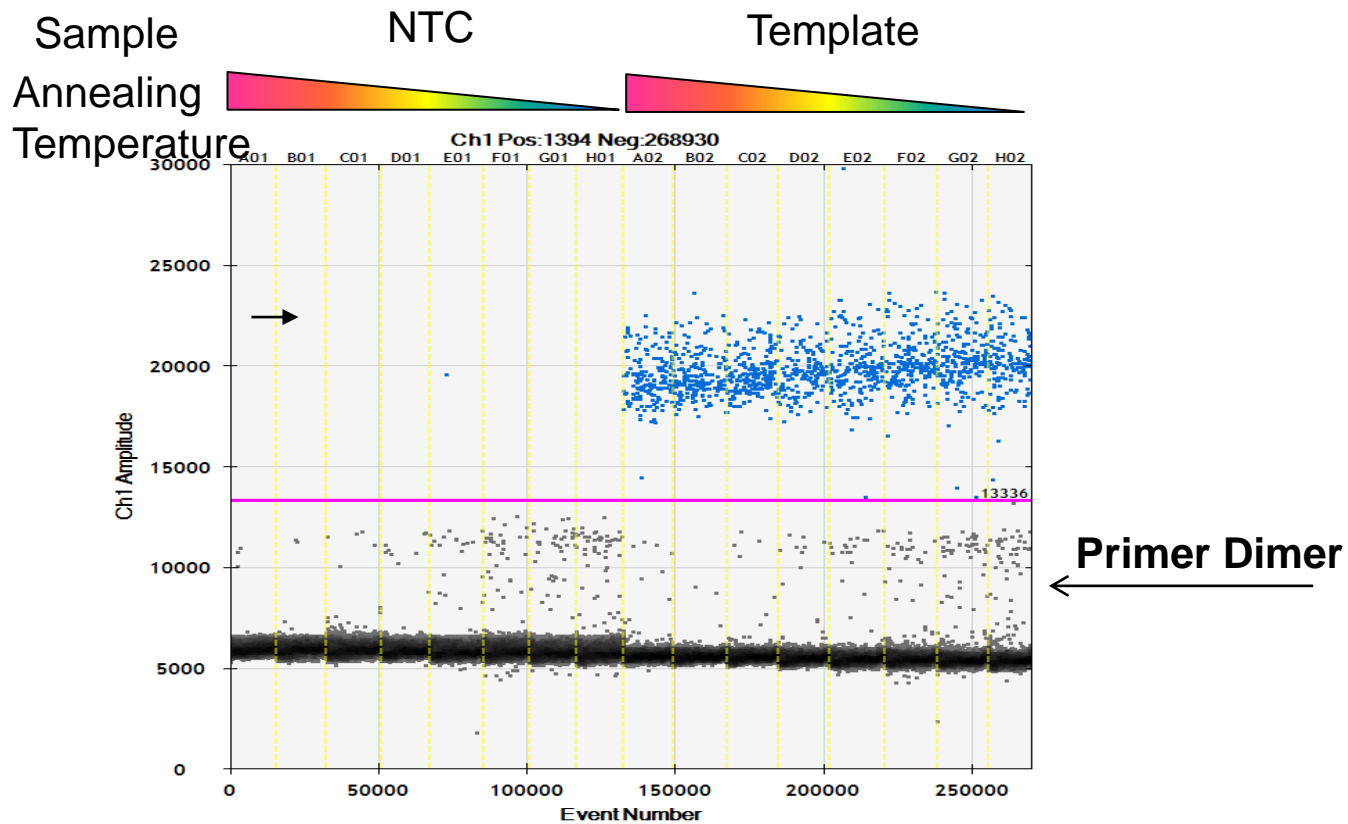


Table 2. Comparison between EvaGreen and TaqMan Mutation Quantification Methods on Control Template DNA

template	source	% mutant	
		EvaGreen	TaqMan
18507	normal diploid DNA	0.01	0.00
Human Male Control	normal diploid DNA	0.00	0.00
HT29	cancer cell line	25.38	25.68
LS411N	cancer cell line	67.48	66.36
168B	patient-benign	0.00	0.00
168M	patient-malignant	25.30	27.13

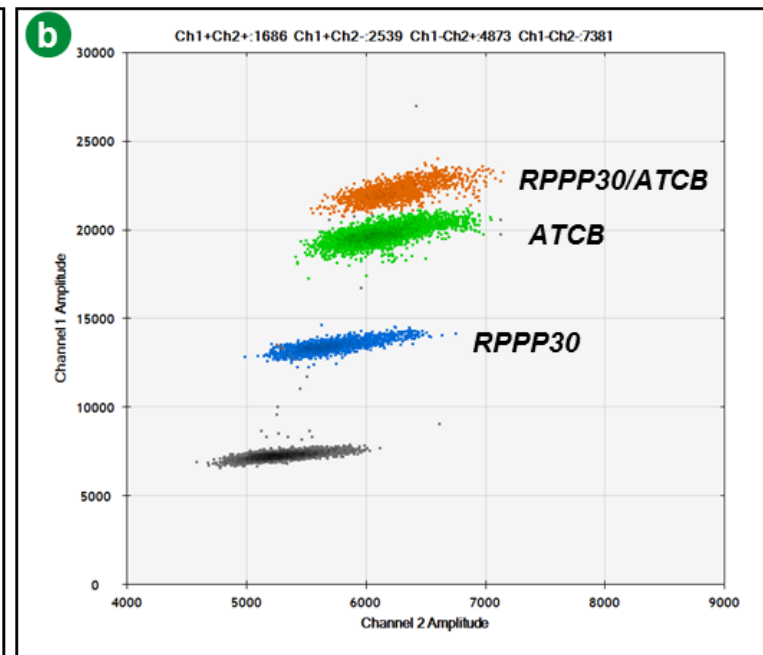
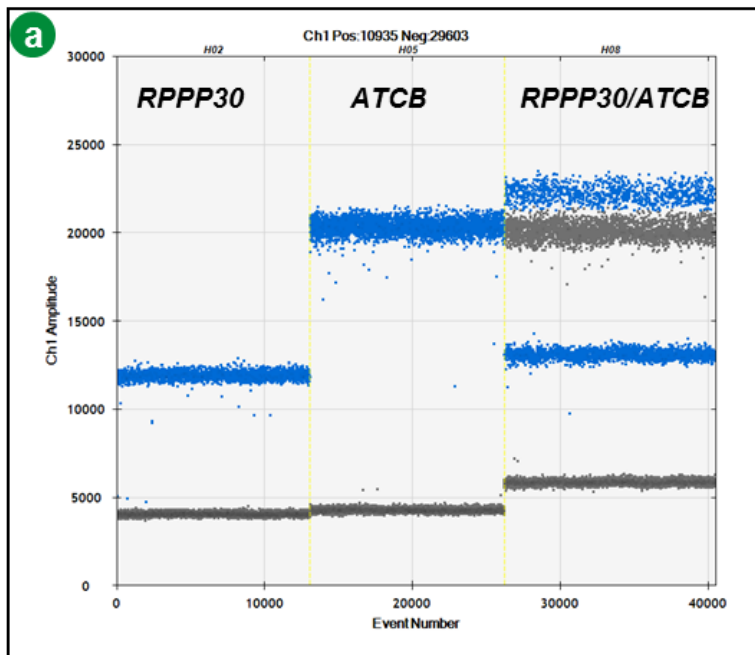
Temperature Gradient: EvaGreen

- Primer dimers can be visualized with EvaGreen chemistry
- Primer dimer frequency increases at lower annealing temperature and are visible in NTCs



Multiplexing with EvaGreen

Multiplexing with EvaGreen by varying amplicon length



RPPP30 amplicon = 62 base pairs
ACTB amplicon = 137 base pairs



Selection Criteria for ddPCR Supermixes



ddPCR Supermix for Probes



ddPCR Supermix for Probes (no dUTP)



QX200 ddPCR EvaGreen Supermix



One-Step RT-ddPCR Kit for Probes

Applications and Considerations

Applications and Considerations	ddPCR Supermix for Probes	ddPCR Supermix for Probes (no dUTP)	QX200 ddPCR EvaGreen Supermix	One-Step RT-ddPCR Kit for Probes
Suitable for UNG decontamination protocols	✓	–	–	–
Compatible with validated PrimePCR ddPCR mutation detection assays	•	✓	–	–
Compatible with validated PrimePCR ddPCR copy number assays	•	✓	–	–
Compatible with PrimePCR gene expression primer assays	–	–	✓	–
ddPCR library quantification kit for Illumina TruSeq	–	✓	–	–
ddPCR library quantification kit for Ion Torrent	–	✓	–	–
Amplification in droplets for downstream sequencing	–	✓	–	–
Double-stranded DNA detection	–	–	✓	–
Absolute quantitation of target RNA molecules	–	–	–	✓

✓ Recommended supermix
• Compatible supermix

PrimePCR™ ddPCR™ Assays

- Designed for digital PCR, fully validated assays
- Can be used on the QX200 ddPCR system
- Universal cycling conditions and primer/probe design strategy
- Universal restriction enzyme strategy for copy number assays
- World class design & manufacturing expertise

Mutation Detection Assays

- Targets for most frequent mutations in COSMIC
- >2500 target with matching wild type assays

BRAF		EGFR		HRAS		KRAS	
V600E	P367R	T790M	L858R	G12V	G13R	G12D	G12S
V600V	F595S	E746 A750	L858M	G12A	G13S	G12A	G13D
S616F	T599I	G719D	H835L	G12R	Q61K	G12V	G13A
D494N	G460G	L747S	E709A	G13V	Q61H	G12C	K5N
K601N	I326T	L861R	P848L	G13C	H27H	G12R	A146T

Copy Number Assays

- All high-value targets in Cancer & Neuro
- >700 target assays + 2 reference assays

APC	AR	ARID1A	ATM	BIRC2	BRCA1	BRCA2	CCND1	CCND2	CCNE1	CDK4	EIF2C1
CDK6	CDKN1A	CRKL	CSMD1	DCUN1D1	DEFB119	EGFR	ERBB3	FGFR1	FGFR2	FOXO1	AP3B1
GAB2	GRB2	HMGA2	IGF1R	IRS2	JUN	KDM6A	KIT	KRAS	MAGI3	MAP2K4	
MDM2	MELK	MET	MTAP	MYB	MYC	MYCN	NCOA3	NCOR1	ORAOV1	PARK2	
PDGFRA	PIK3CA	PPP2R1A	PSEN1	PTEN	RB1	REL	RPS6KB1	SHH	SKP2	SLIT2	
		SMAD4	TERT	TSC1	TSC2	WHSC1L1	WISP1	YAP1			



You can use our look up tool to see if we have it or not:

www.bio-rad.com/primepcr_lookup

Publication Database

BIO-RAD

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More than 8300 published studies have described research breakthroughs using Droplet Digital™ PCR technology

8391

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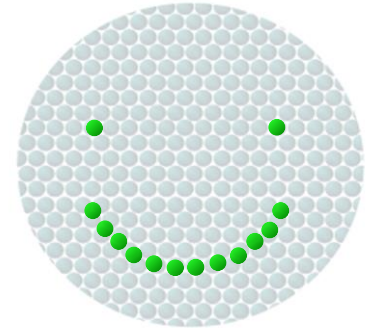
Reset

Sort By Alphabetical-Asc

< 1 2 3 4 ... 420 >

Best Reasons to use ddPCR

- **Sensitivity**
 - 10-1000x fold improvement over qPCR
 - Works with FFPE blood and tissues samples
- **Absolute Quantification**
 - Quantify gene expression more precisely
 - No Standard Curve
- **Precision**
 - Measure more subtle differences in expression or mutation
 - Detect structural variants in cancers
- **High Throughput**
 - Maximum 4 x 96 per day
 - Save \$ and Time



Q&A



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