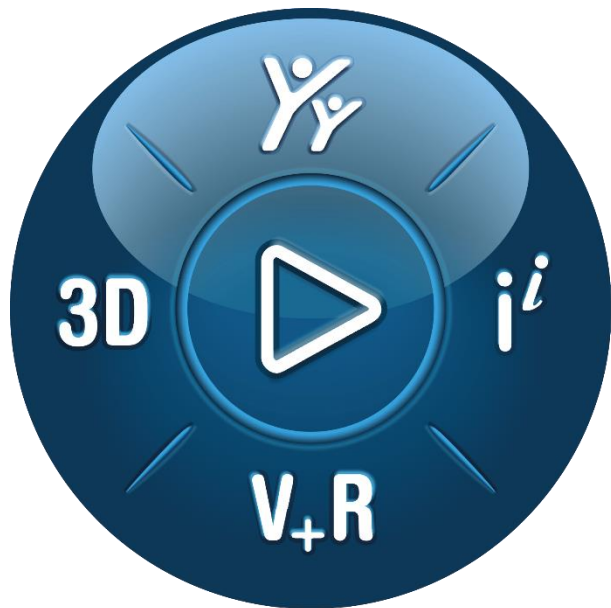


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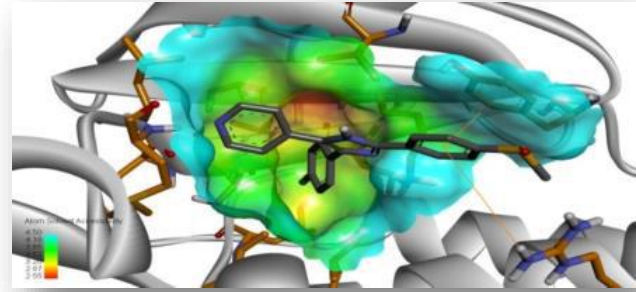
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Simulation

- Assume.....that we have a molecule on the computer screen
- It could have been
 - Built from scratch
 - Prepared from a PDB file
 - Read in from a prepared file
- But...How do you know you have a reasonable structure?



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Simulation

- Molecular Mechanism (MM), Molecular Dynamic (MD) simulation
 - Assignment of the force field
 - CHARMM, charmm19/22/27/36, CHARMM Polar H, MMFF, CFF
 - Additional solvent model
 - implicit/explicit water molecules, Implicit Membrane
 - Additional constraints
 - Fix, Dihedral, Harmonic, Distance
- Energy calculation
 - Time point energy calculation
 - Structural optimization
 - Structural optimization using QM (DFT)
 - MD (CHARMM, NAMD)
- Analyze trajectory
 - Plot of various properties – distance, angle, interaction energy, PCA, RDF, Radius of Gyration

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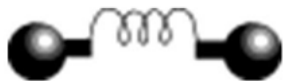
Molecular Mechanism (MM)

- Based on the classical molecular mechanics
 - Energy calculations
 - Energy minimizations
 - Molecular dynamics
- Widely used in
 - Structure-based ligand design
 - Structure generation from NMR experiments
 - Protein engineering
 - Substrate recognition studies

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Force Field

“The molecule is considered to be a collection of atoms held together by simple elastic or harmonic forces...”



$$E = k_b (r - r_0)^2$$

Adopted from computer software applications in chemistry, Peter C. Jurs, 1986

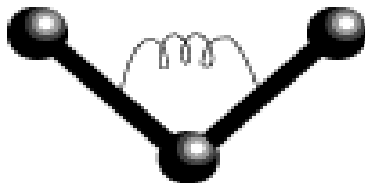
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Force Field

“The molecule is considered to be a collection of atoms held together by simple elastic or harmonic forces. The forces are defined in terms of **potential energy** described by the **internal coordinates** of the molecule...”



$$E = k_b(r - r_0)^2$$



$$E = k_\theta(\theta - \theta_0)^2$$



Adopted from Computer Software Applications in Chemistry, Peter C. Jurs, 1986

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Force Field Energy Expression

- General form

$$E = E_{bond} + E_{angle} + E_{torsion} + E_{impr} + E_{nonbond} \\ (E_{vdw} + E_{elecs}) \\ + E_{other}$$

- Force fields are generally differ in:
 - Function expression of each term
 - Parameter calculation method
 - Function parameters
 - Cross terms

$$E_{pot} = \sum k_b(r - r_0)^2 + \sum K_\theta(\theta - \theta_0)^2 + \sum |k_\phi| - k_\phi \cos(n\phi) + \sum k_\chi(\chi - \chi_0)^2 \\ + \sum \frac{q_i q_j}{\epsilon 4\pi \epsilon_0 r_{ij}} + \sum \left(\frac{A_{ij}}{r_{ij}^{12}} - \frac{B_{ij}}{r_{ij}^6} \right) sw(r_{ij}^2, r_{on}^2, r_{off}^2) + E_{constraint} + E_{user}$$

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CHARMm force field

- 2 official version

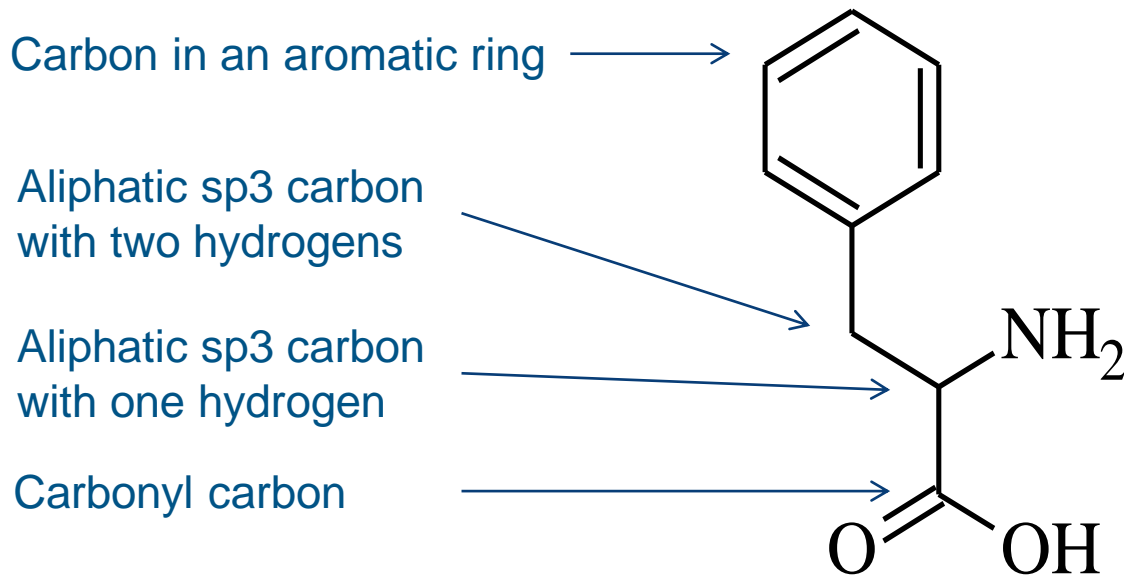


- Harvard academic version (Professor Martin Karplus, the 2013 Nobel laureate in Chemistry)
- DS integration (Graphical interface)
 - Integration with other programs that use the CHARMM engine (CDOCKER, MCSS, ZDOCK)
 - GBSA / IM solvation model

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Force Field Parameters

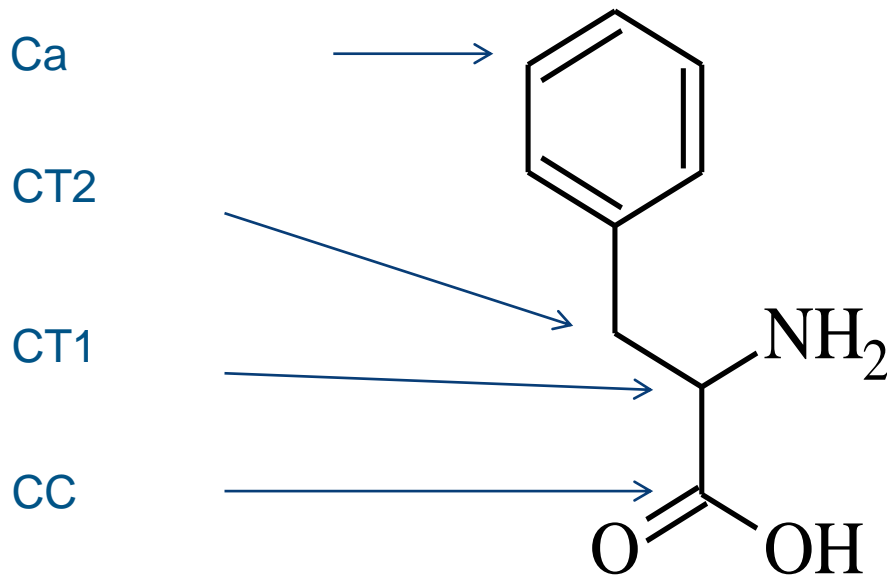
- Atoms have different environments



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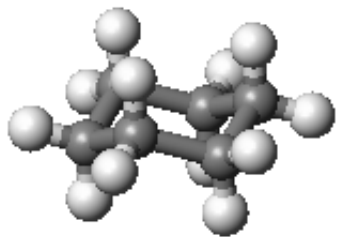
Force Field Parameters

- ...which yield different parameters.

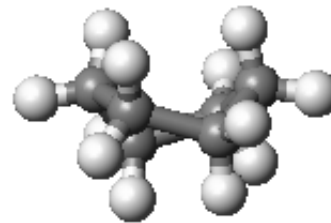


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Molecular Mechanics



12.3 kcal/mol



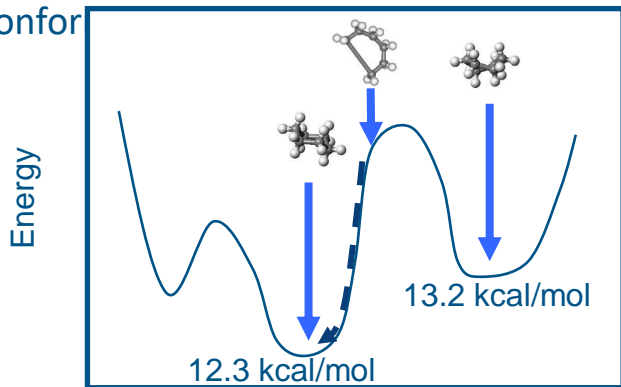
13.2 kcal/mol

It is the difference in energy that matters!

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Energy Minimization

- If lower energy conformations are more favorable, we may want to find them.
- Could perform an energy minimization
 - A mathematical technique to find a local minimum for the energy expression shown earlier
 - Adjust Cartesian coordinates to attain the more favored conformation
- When to perform an energy minimization?
 - Clean a structure that you build
 - Prepare a structure from a PDB file
 - Relax a modified structure
 - Relax a docked protein-ligand complex

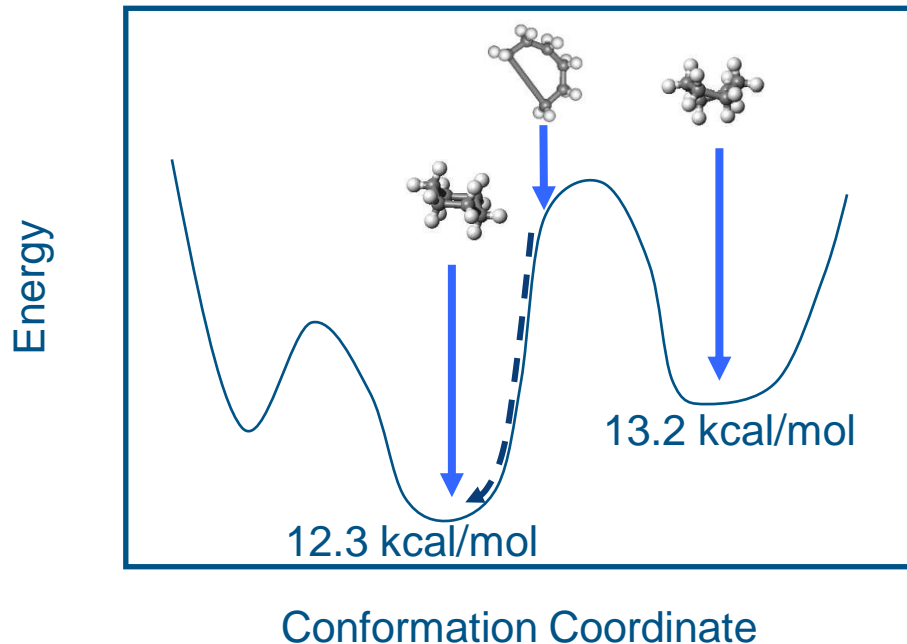


Conformation Coordinate
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Molecular Dynamics

- Global vs Local minima

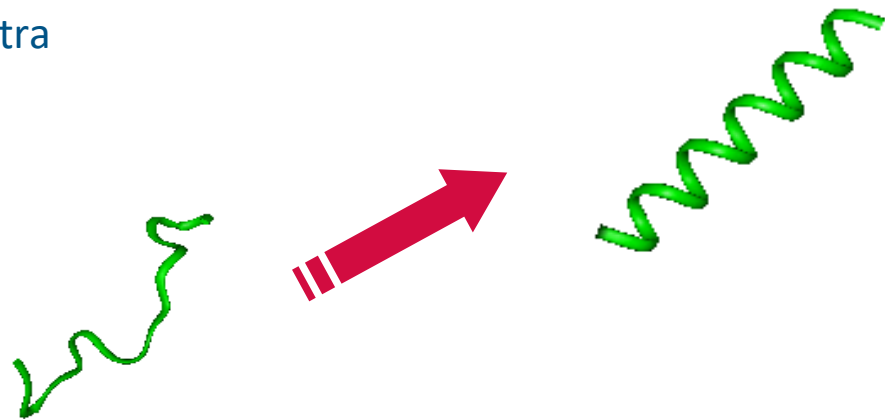
How to explore other minima or other areas of the potential energy surface?



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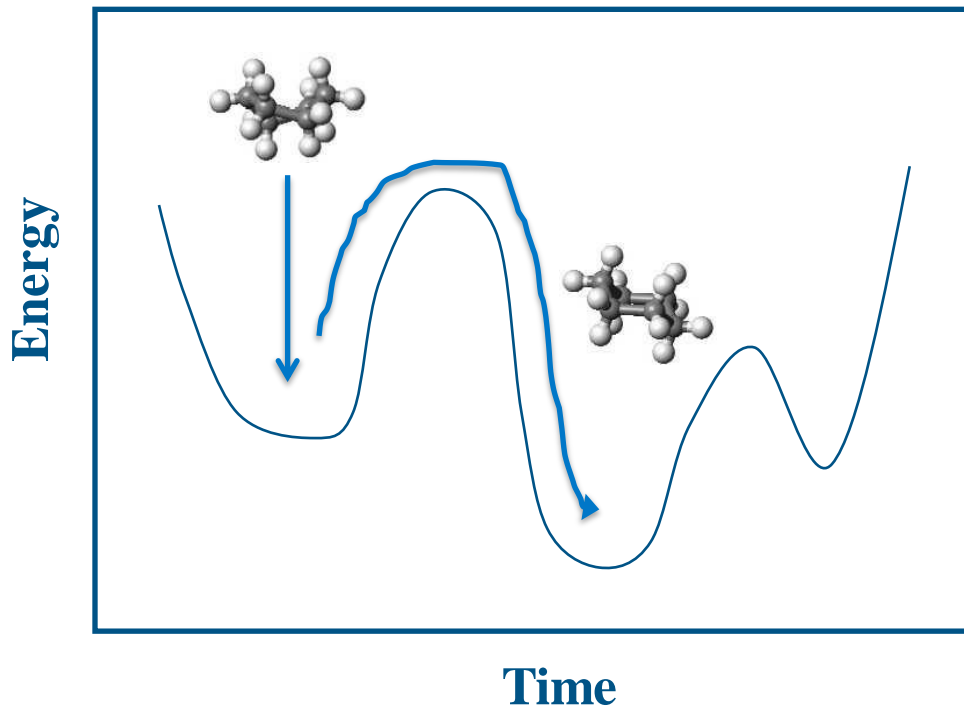
Use of Molecular Dynamics

- Conformational changes of molecule
 - Simple vibrations that produce spectra
 - Breathing of DNA
 - Hinge bending
 - Allosteric transitions
 - Protein folding
 - Refinement of docking results
 - Induced fit
- Free energy changes
 - Calculation to binding energies
- Determination of thermodynamic properties
 - Enthalpy and entropy changes



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Example for MD simulation



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Temperature and Velocities

- From kinetic theory, temperature is proportional to the kinetic energy of the particles.

$$KE = \frac{3}{2}kT$$

- From physics, kinetic energy is proportional to velocity.

$$KE = \frac{3}{2}mv^2$$

- Therefore, temperature is proportional to the velocity of the atoms.

$$T = \frac{mv^2}{k}$$

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Temperature and Dynamics

At a higher temperature...

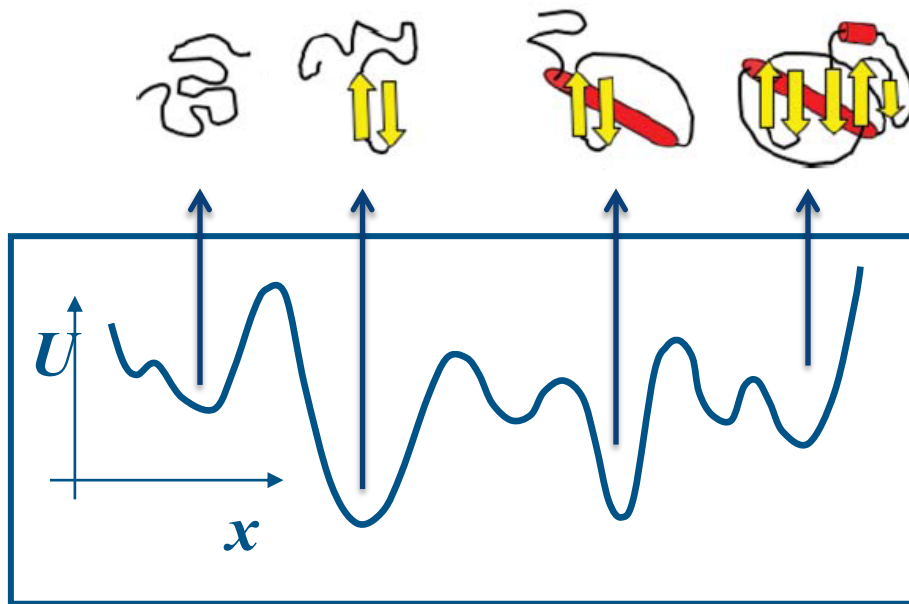
Atoms move more...

Cover more conformational space!

- High temperature dynamics can be used to overcome barriers to explore new local minima
- Effective for reasonable length peptides and other oligomers (12 residues or less)
- Must be aware of
 - Possible loss of secondary structure
 - Trans-cis isomerization of peptide bonds

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Energy landscape: searching the possible conformations

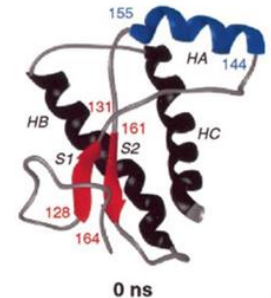
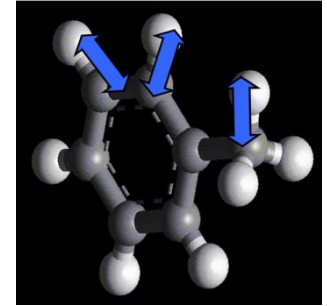


Local minimum

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Time scale of Biological events

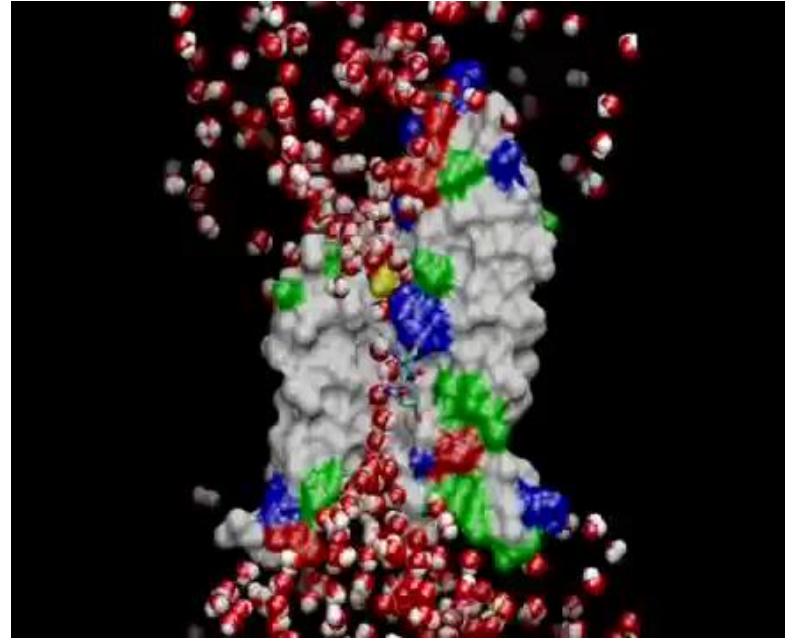
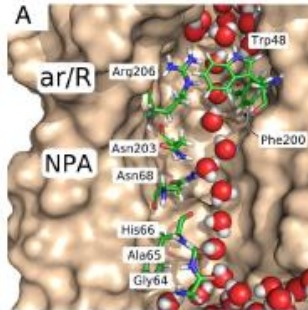
Motion	Time scale (Sec)
Bond stretching	10^{-14} to 10^{-13}
Elastic vibrations	10^{-12} to 10^{-11}
Rotations of surface side chains	10^{-11} to 10^{-10}
Hinge bending	10^{-11} to 10^{-7}
Rotation of buried side chains	10^{-4} to 1 sec
Allosteric transitions	10^{-5} to 1 sec
Local denaturations	10^{-5} to 10 sec



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Usage of MD simulation

- Example: Aquaporin
- Aquaporins form a family of pore proteins that facilitate the efficient and selective flux of small solutes across biological membranes



Mechanism of selectivity in aquaporins and aquaglyceroporins. Hub JS, de Groot BL. Proc Natl Acad Sci U S A, 2008

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Usage of MD simulation

- Example: GB1 folding
- IgG-binding protein G

This movie supplements the article:
Folding pathway of the B1 domain of protein G
explored by a multiscale modeling.
Kmiecik and Kolinski, Biophys J, 2007

Ab initio simulation of protein GB1 folding

-- Laboratory of Theory of Biopolymers
-- <www.biocomp.chem.uw.edu.pl>

Folding pathway of the b1 domain of protein G explored by multiscale modeling. Kmiecik S, Kolinski A. Biophys J. 2008

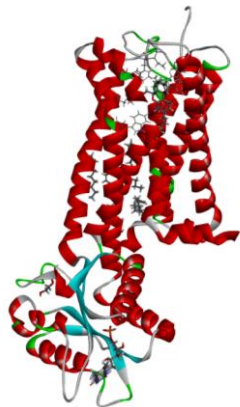
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Molecular Dynamics in DS

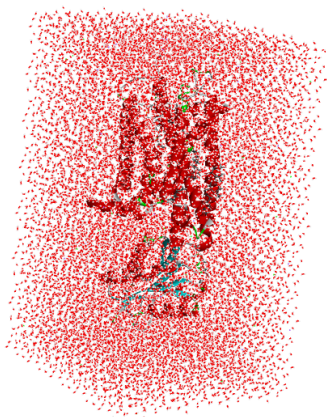
- Before Run MD
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 - Explicit Membrane-Based Solvation
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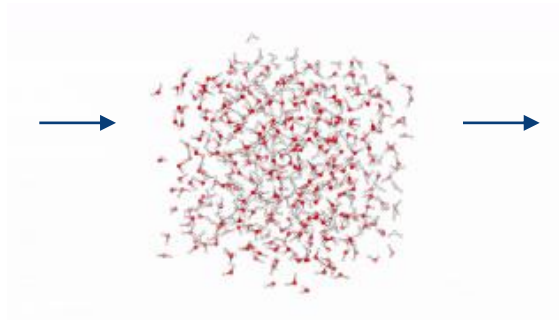
MD steps



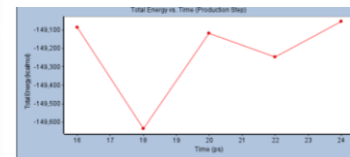
Build Model



Solvation



Run MD



Analysis

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Build Model

- Model for MD simulation can be obtain by...
 - Protein Data Bank
 - Alphafold
 - Theoretical Model
 - Homology Modeling
 - Docking
 - Others...

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Solvation

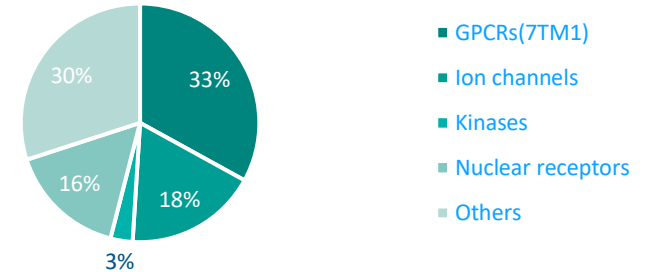
- Implicit solvent model
 - Distance –Dependent Dielectrics
 - Generalized Born
 - GBMV, GBSW, GBIM
- Explicit solvent model
 - Cell shape
 - Periodic Boundary Distance
 - Counterion

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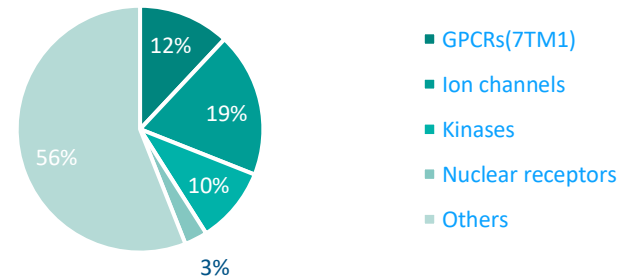
Explicit Membrane-Based MD Simulations

- Membrane proteins account for approximately 1/3 of the human proteome and account for ~60% of pharmaceutical targets⁺
- These membrane proteins include a large number of signaling receptors, transporters, ion channels and enzymes that are vital to cellular regulation, metabolism and homeostasis, and are high-priority, pharmaceutically relevant research
- CHARMM has the best and most consistent set of lipid parameters, including sterols, unusual bacterial, mitochondrial, and endosomal lipids, and a consistent set of protein parameters, to model complex membranes and protein membrane interactions⁺

Proportion of human protein drug targets in major families

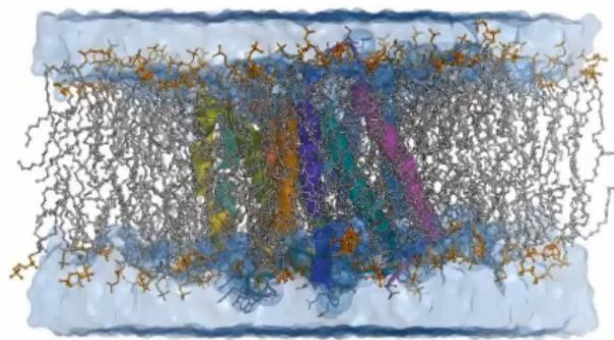
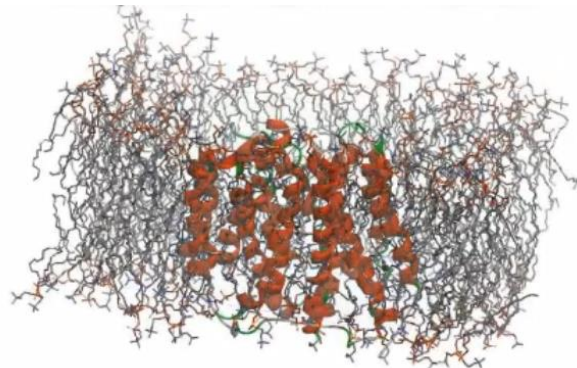


Proportion of small-molecule drugs that target major families



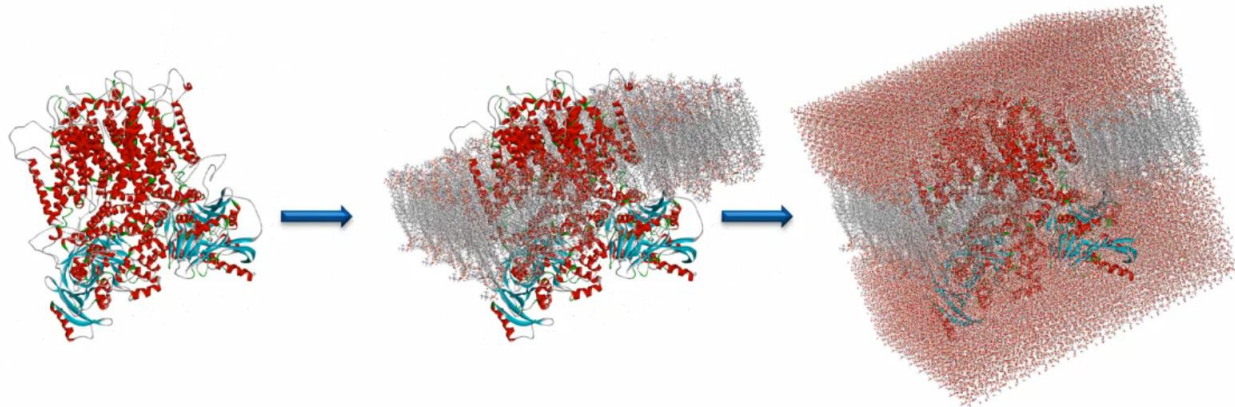
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- CHARMM has the best and most consistent set of lipid parameters, including sterols, unusual bacterial, mitochondrial, and endosomal lipids, and a consistent set of protein parameters, to model complex membranes and protein membrane interactions⁺



Explicit Membrane-Based MD Simulations

- New protocol adds an explicit bilayer of lipids, water and counterions to a transmembrane protein and allows the option to equilibrate the system, prior to further simulation
- Supports a number of prepared equilibrated homogenous membranes, as well as complex custom membranes (e.g. <http://www.charm-gui.org>)
 - POPC, POPE, DPPC, DMPC, and DLPC



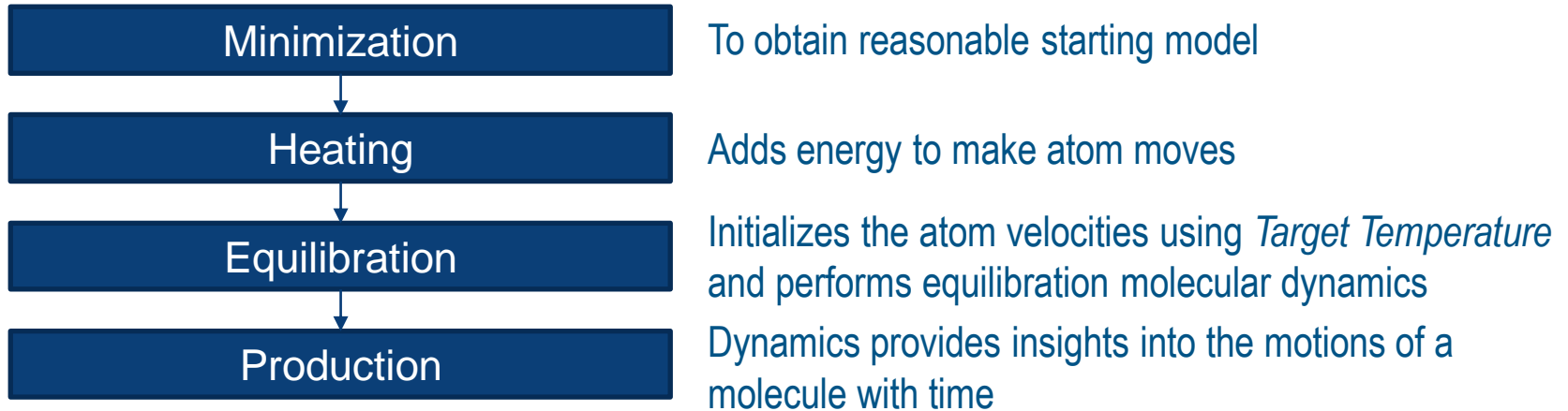
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Standard Dynamic Cascade (SDC)

- General MD in DS
- Based on CHARMM Forcefield



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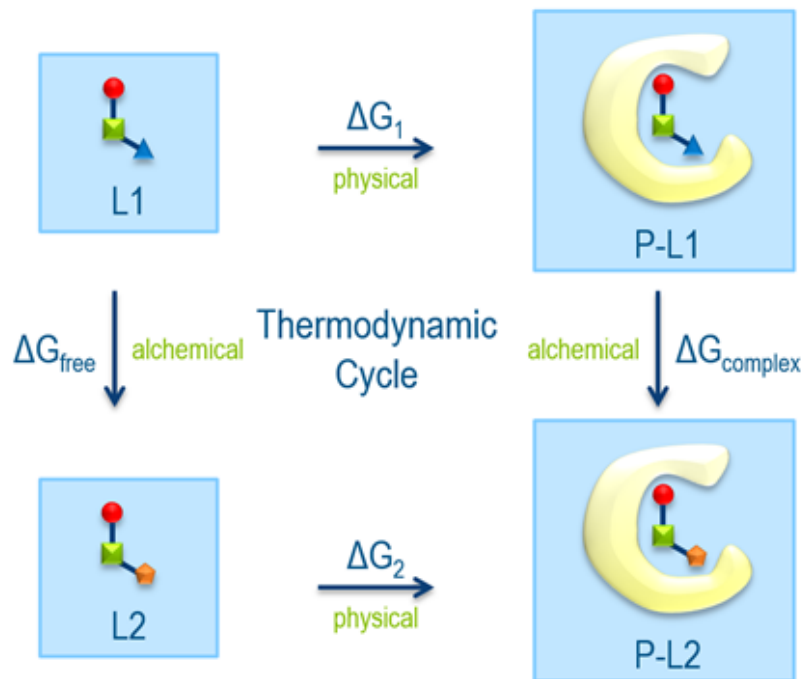
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Free Energy Perturbation

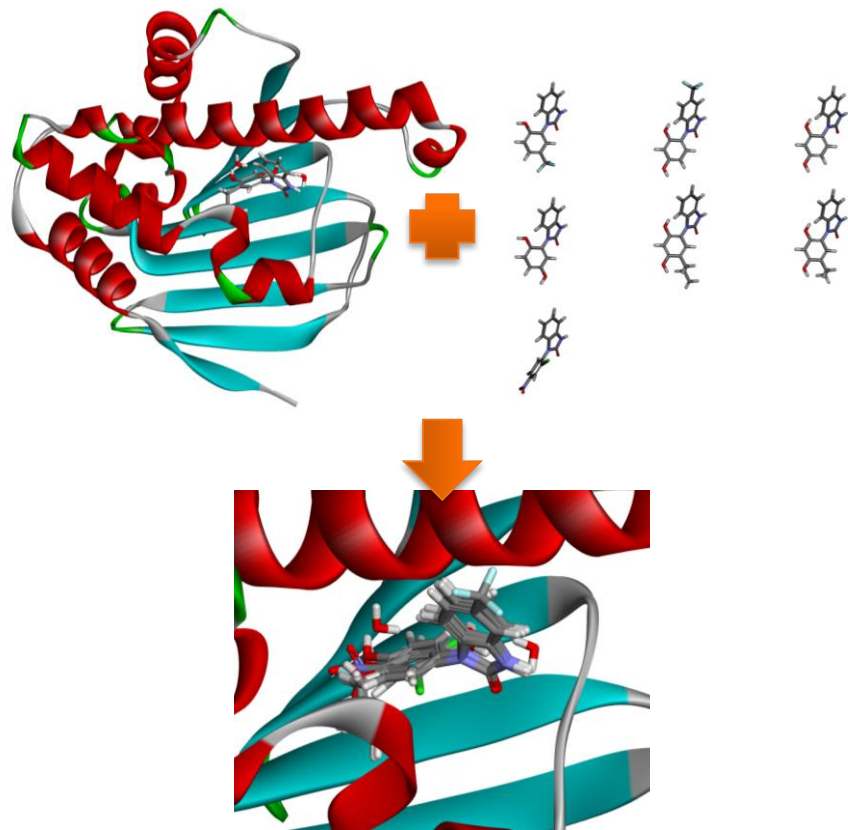
- The ability to understand and quantify the strength of interactions between a ligand and protein is essential in drug discovery projects to discern how changes to a ligand might alter affinity.
- Alchemical free energy perturbation (FEP) is based on a thermodynamic cycle.



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Generate Analog Conformation

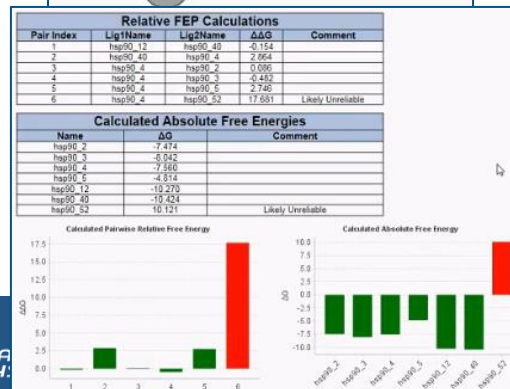
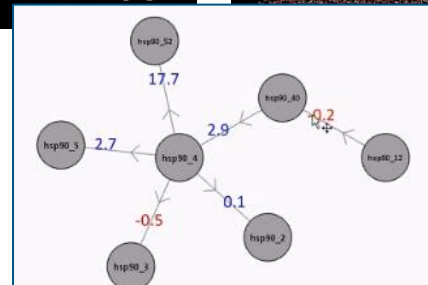
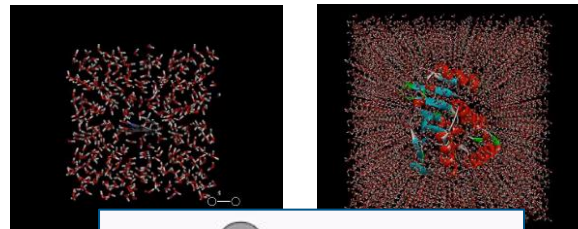
- Receptor
 - Receptor should be prepared well before run FEP
- Docked Lead
 - Used as reference to align analogues to the binding site
- Analog
 - Superimpose is recommended before run FEP



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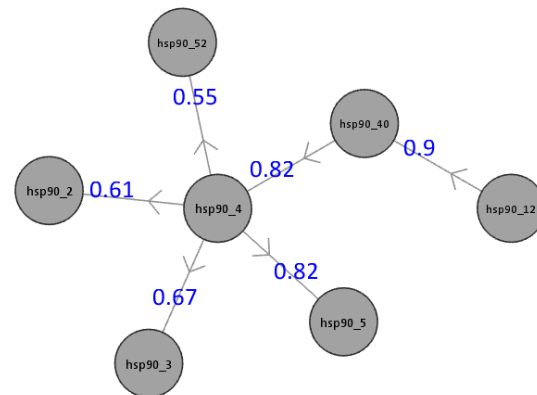
Predict Relative Free Energy of Binding

- Apply FEP to a congeneric analog series in four steps using NAMD
 - Use either docking results, or *in-situ* analog series as input
 - Optional: Type analogs with charmm36/GCenFF using MATCH engine
- Step 1: Generate a ligand pairs network map
 - Builds a pairs network using maximal common substructures
- Step 2: Set up relative FEP calculations
 - Generates explicit solvent models for each ligand pair and protein-ligand pair complex in the network map
- Step 3: Run free energy perturbation calculations
 - Perform FEP simulations on each solvated ligand pair model
- Step 4: Collate the simulation results
 - Summarises both relative free energy calculations and absolute free energies (if a free energy value is available for the reference lead)



Generate a ligand pairs network map

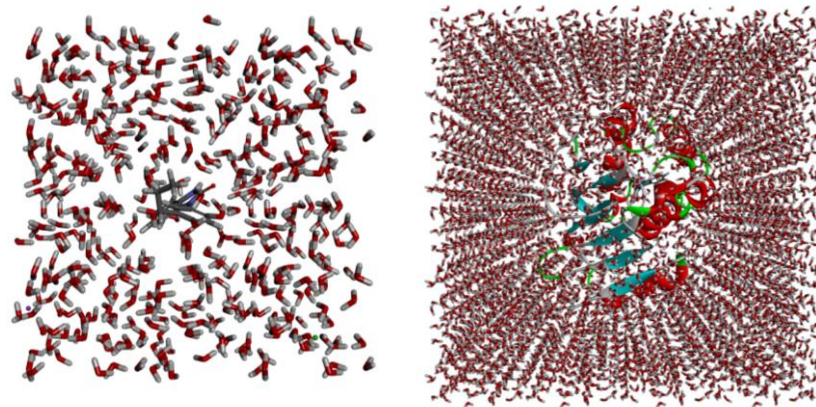
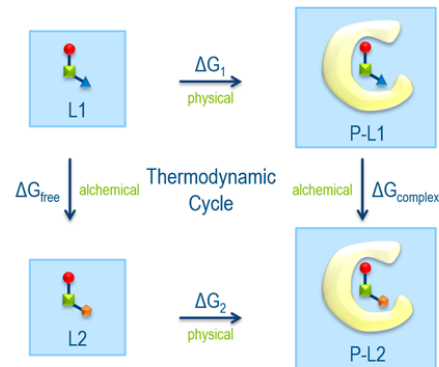
- Generates a set of optimal pairs based on chemical and topological similarity in order to maximize the total similarity of a network spanning the entire set.
- Similarity matrix using a Maximal Common Substructure (MCS) method.
- The similarity between any two ligands L1 and L2 is computed as $\exp[-\text{Beta} \cdot (nL1 + nL2 - 2 \cdot nMCS)]$



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Set up relative FEP calculations

- Generates explicit solvent models for each ligand pair and protein-ligand pair complex in the network map
- The calculation is set up to transform the first ligand in each pair into the second. For each pair, a merged dual topology molecule is generated based on the identified common and different set of atoms in the topologies and nonbonded parameters.

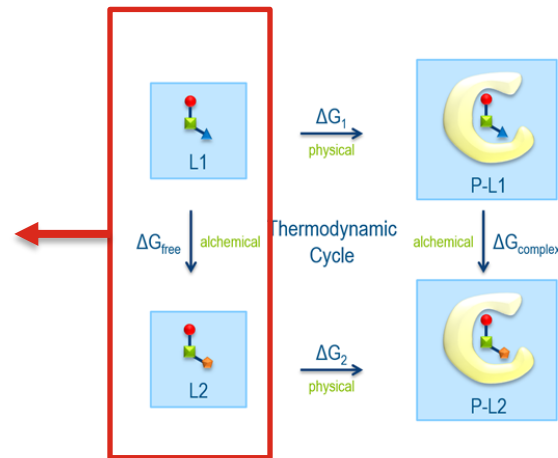


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Run free energy perturbation calculations

- Runs alchemical Free Energy Perturbation (FEP) dynamics simulations using **NAMD**
- Solvated Ligand and Solvated Complexes are run separately.

FEP Simulation Results							
Name	Lambda Range	Calc ΔG	StDev	SimulationType	FEP Report	Simulation Trajectory	Simulation Trajectory Without Solvent
hsp90_12--hsp90_40	0-1	1.02	0.04	Ligand	FEP Report	Trajectory	Trajectory
hsp90_40--hsp90_4	0-1	0.90	0.05	Ligand	FEP Report	Trajectory	Trajectory
hsp90_4--hsp90_5	0-1	-10.01	0.09	Ligand	FEP Report	Trajectory	Trajectory
hsp90_4--hsp90_3	0-1	4.04	0.02	Ligand	FEP Report	Trajectory	Trajectory
hsp90_4--hsp90_2	0-1	2.46	0.12	Ligand	FEP Report	Trajectory	Trajectory
hsp90_4--hsp90_52	0-1	20.30	0.63	Ligand	FEP Report	Trajectory	Trajectory



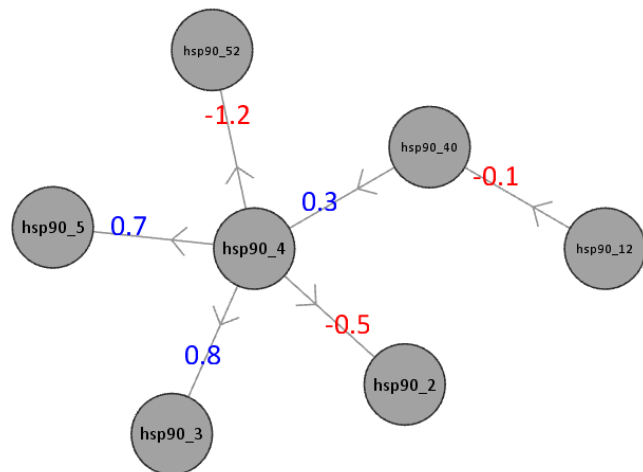
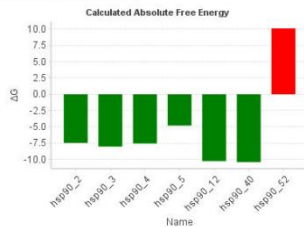
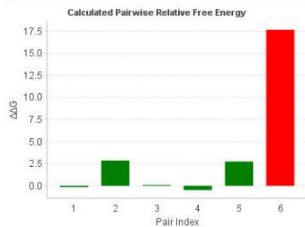
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Collate the simulation results

- The *Collate FEP Simulation Results* protocol also generates a network plot (pictured) that graphically presents the $\Delta\Delta G$ values of the ligand pairs. The edges are labeled and color-coded with the relative free energy of the ligand pair, and the arrow head indicates the direction of the alchemical change.

Relative FEP Calculations							
Pair Index	Lig1Name	Lig2Name	$\Delta\Delta G$	StdDev	Calc. ΔG (Ligand)	Calc. ΔG (Complex)	Comment
1	hsp90_12	hsp90_40	-0.154	0.101	7.16	7.01	
2	hsp90_40	hsp90_4	2.884	0.054	-9.39	-6.53	
3	hsp90_4	hsp90_2	0.096	0.176	20.03	20.11	
4	hsp90_4	hsp90_3	-0.482	0.083	13.85	13.37	
5	hsp90_4	hsp90_5	2.746	0.084	-0.64	2.11	
6	hsp90_4	hsp90_52	17.681	0.503	1.54	19.22	Likely Unreliable

Calculated Absolute Free Energies		
Name	ΔG	Comment
hsp90_2	-7.474	
hsp90_3	-8.042	
hsp90_4	-7.560	
hsp90_5	-4.814	
hsp90_12	-10.270	
hsp90_40	-10.424	
hsp90_52	10.121	Likely Unreliable



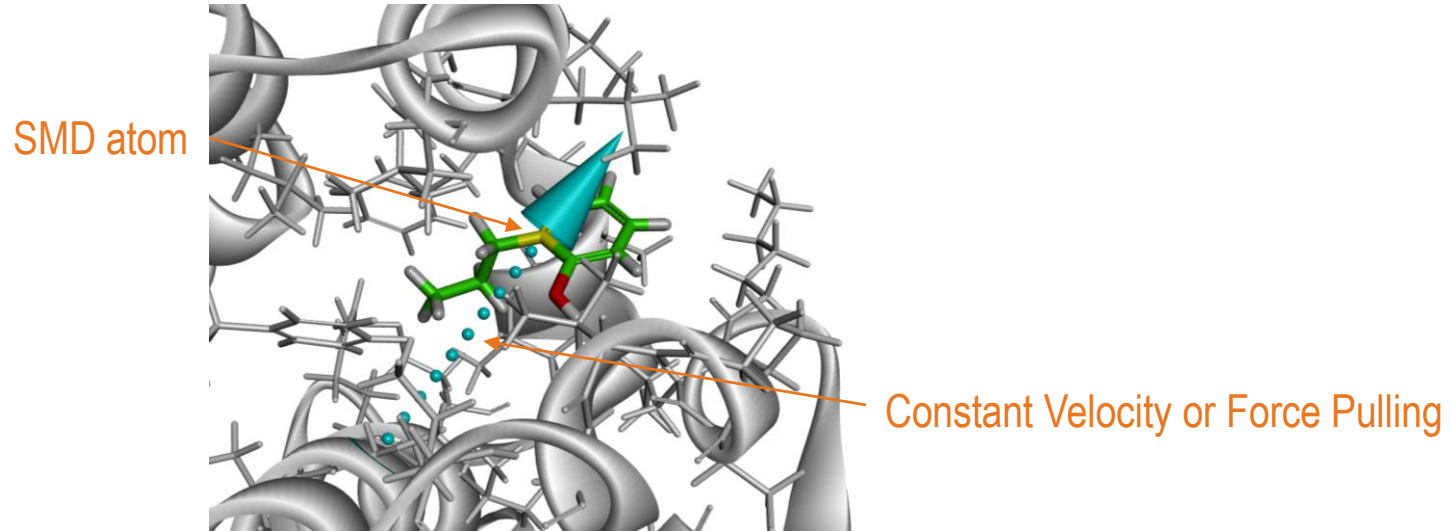
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Steered Molecular Dynamics

- The basic idea behind any SMD simulation is to apply an external force to one or more atoms, which we refer to as SMD atoms.



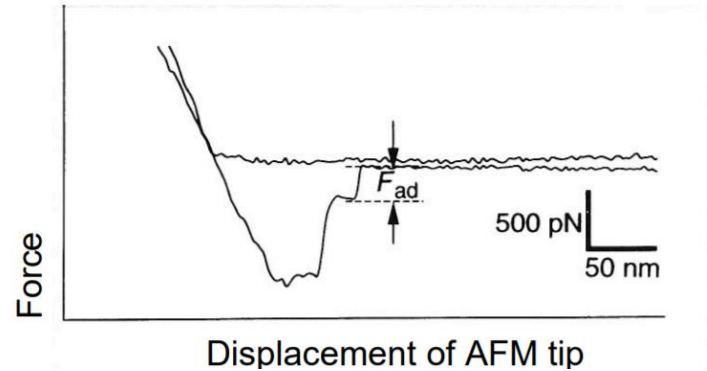
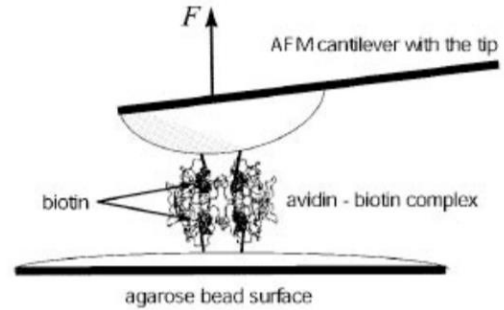
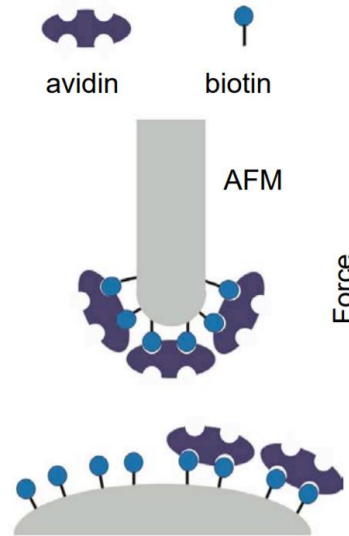
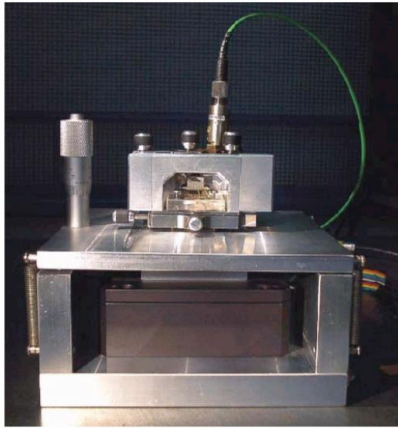
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Why SMD

- Accelerates processes to simulation time scales (ns)
- Yields explanations of biopolymer mechanics
- Complements Atomic Force Microscopy
- Finds underlying unbinding potentials
- Generates and tests Hypotheses

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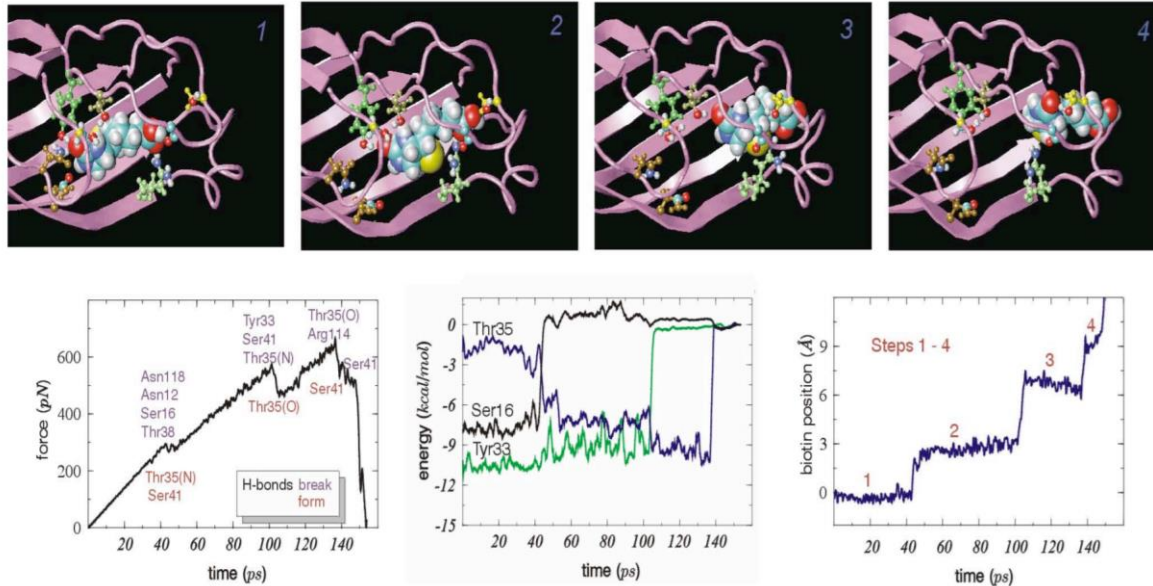
Atomic Force Microscopy Experiments of Ligand Unbinding



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SMD of Biotin Unbinding: What We Learned

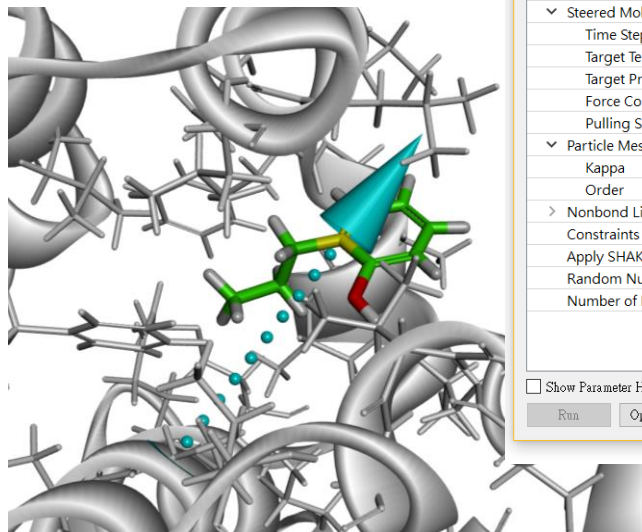
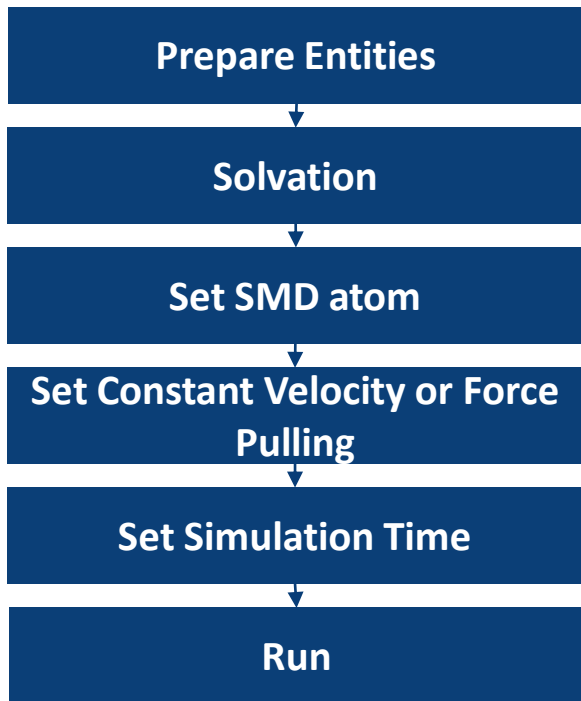
- biotin slips out in steps, guided by amino acid side groups, water molecules act as lubricant, MD overestimates extrusion force



Israilev et al., *Biophys. J.*, 72, 1568-1581 (1997)

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General Steps in MD



Steered Molecular Dynamics

Parameter Name	Parameter Value
Input Typed Molecule	1HK0:1HK0
Simulation Time (ps)	1000
Save Results Interval (ps)	5
Pulling Atom Pair	
Advanced	
Steered Molecular Dyn...	
Time Step (fs)	2
Target Temperature	300
Target Pressure	1.0
Force Constant	500.0
Pulling Speed	0.02
Particle Mesh Ewald	
Kappa	0.34
Order	4
> Nonbond List Radius	14.0
Constraints	
Apply SHAKE Constraint	True
Random Number Seed	
Number of Processors	1

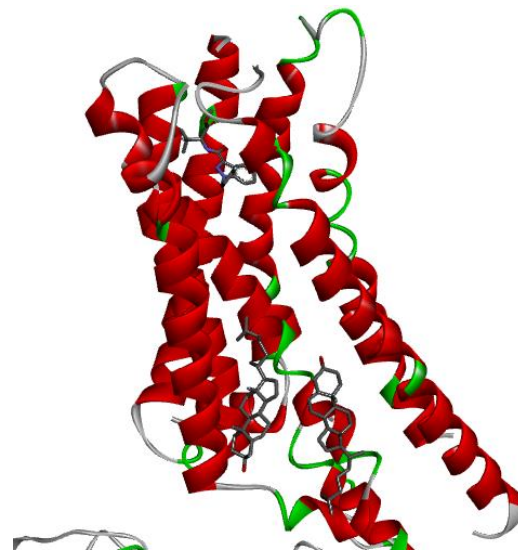
Show Parameter Help

Run Options Cancel Help

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Hands on

- Cannabinoid Receptor 1-G Protein Complex
- methyl N-{1-[(4-fluorophenyl)methyl]-1H-indazole-3-carbonyl}-3-methyl-L-valinate
- Predict the unbounding pathway and binding free energy of the *in situ* ligand



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Molecular Dynamics in DS

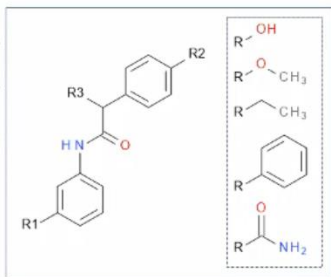
- Before Run MD
 - Solvation
 - Explicit Membrane-Based Solvation
- MD in DS
 - Standard Dynamic Cascade (SDC)
 - NAMD
 - Free Energy Perturbation
 - Steered Molecular Dynamics (SMD)
 - Multi-Site Lambda Dynamics (MSLD)

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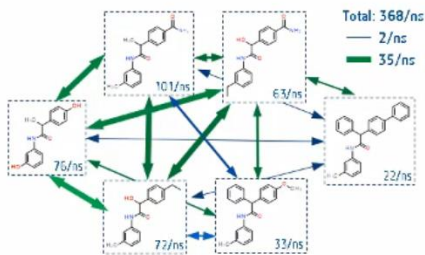
Free Energy: Multi-Site Lambda Dynamics⁺

- Explore large congeneric chemistry space in early/ stage Lead Optimization in a way that FEP can't
 - Based on Lambda Dynamics with more than two decades of development and refinement, unique to CHARMM
 - Multiple ligands in a single simulation, mimicking a competitive binding assay
 - Up to 20 times more efficient than FEP
 - User-friendly and easy to setup, run and analyze

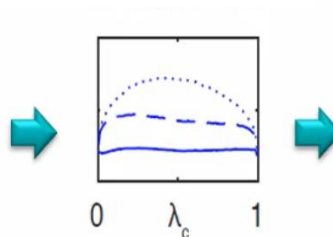
Simulate a competitive binding assay
of a combinatorial library of ligands



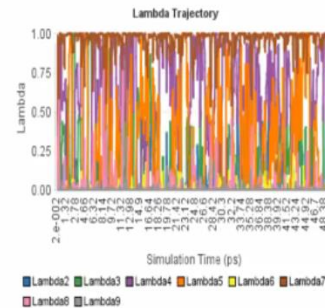
E.g., 5x5x5=125 analogs



Flattening free energy
landscape enhances sampling
and leads to efficient
calculation of binding affinities



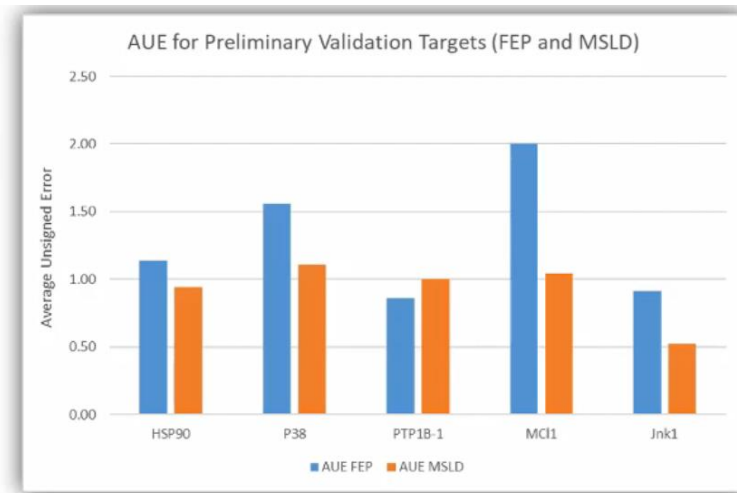
Obtain ligand affinities
from occupancy ratios



MSLD: Preliminary Validation

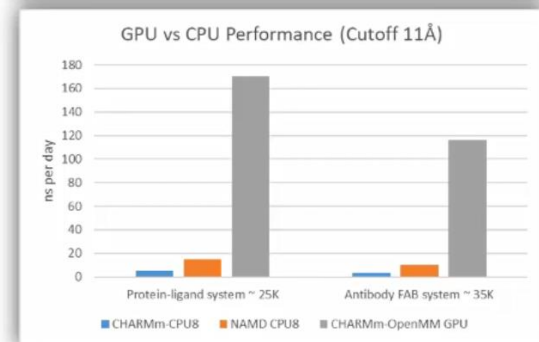
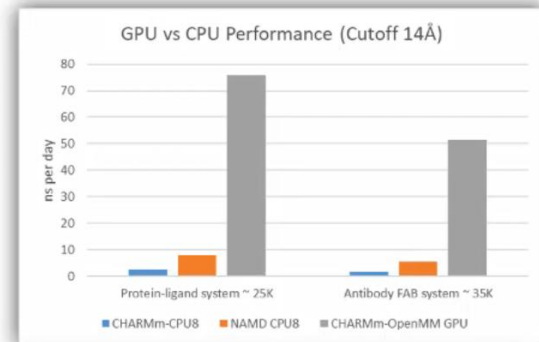
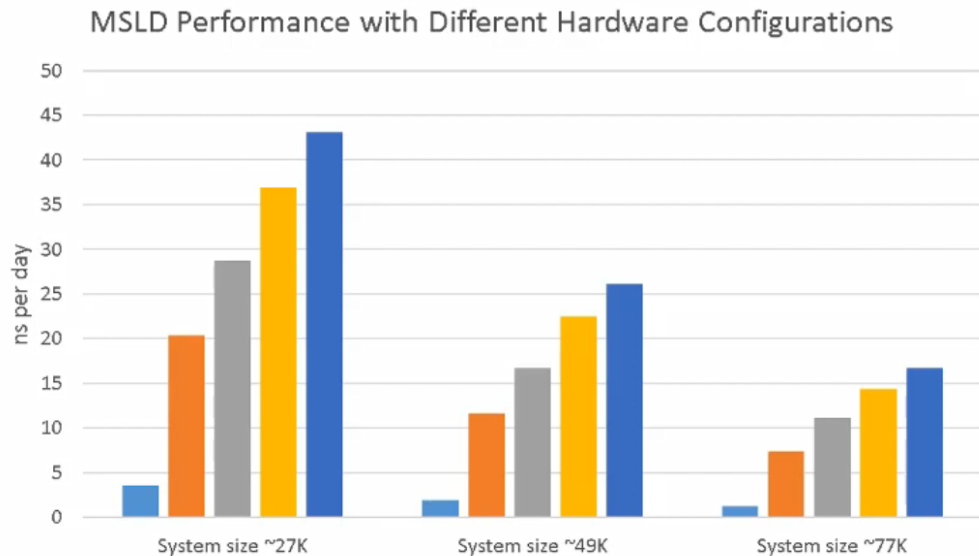
- MSLD-domain datasets with experimental data for validation are difficult to find
 - Permutation of multiple sites on a common core
 - Large dynamic range of relative binding free energy for realistic ranking challenge
- Preliminary validation
 - 5 protein targets, 48 ligands
 - Compare to existing FEP results (up to 32ns sampling per pair)
- MSLD validation parameters
 - Setup systems in cubic boxes and run on domdec-GPU (no restraints)
 - Bias Optimization: ~100 iterations 100ps length each
 - 30ns of production, in triplicate (no replica exchange)
 - Compute simple average of ΔG_{solv} and ΔG_{prot} to calculate $\langle \Delta G \rangle$ and σ
- MSLD average unsigned error of predicted affinities less than FEP
 - Approximately 1 kcal/mol

Set	Sites	nLigands	Range (kcal/mol)
HSP90	2	9 (6x5)	4.3
P38	1	6	1.5
PTP1B-1	1	14	2.8
MCI1	1	8	2.2
Jnk1	1	11	2.1



Multi-Site Lambda Dynamics - MSLD

- In DS2019 CHARMM-OpenMM GPU on Linux accelerated MD production
- In DS2020 MSLD is available through CHARMM-DOMDEC on Linux for both CPU and GPU



Q&A

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